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# 1 **Weight of evidence tools in the prediction of acute fish toxicity**

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## 6 7 **Abstract**

8 Acute fish toxicity (AFT) is a key endpoint in nearly all regulatory implementations of  
9 environmental hazard assessments of chemicals globally. While it is an early tier assay, the AFT  
10 assay is complex and utilizes many juvenile fish each year for the registration and assessment of  
11 chemicals. Thus, it is imperative to seek animal alternative approaches to replace or reduce  
12 animal use for environmental hazard assessments. A Bayesian Network (BN) model has been  
13 developed that brings together a suite of lines of evidence (LoEs) to produce a probabilistic  
14 estimate of acute fish toxicity without the testing of additional juvenile fish. Lines of evidence  
15 include chemical descriptors, mode of action (MOA) assignment, knowledge of algal and  
16 daphnid acute toxicity, and animal alternative assays such as fish embryo tests and *in vitro* fish  
17 assays (e.g., gill cytotoxicity). The effort also includes retrieval, assessment and curation of  
18 quality acute fish toxicity data as these act as the baseline of comparison to model outputs. An  
19 ideal outcome of this effort would be to have global applicability, acceptance and uptake,  
20 relevance for predominant fish species used in chemical assessments, be expandable to allow  
21 incorporation of future knowledge and data be publicly available. The BN model can be  
22 conceived as having incorporated principles of tiered assessment and whose outcomes will be  
23 directed by the available evidence in combination with prior information. We demonstrate that  
24 as additional evidence is included into prediction of a given chemical's ecotoxicity profile, both  
25 the accuracy and the precision of the predicted AFT can increase. Ultimately an improved  
26 environmental hazard assessment will be achieved.

## 27 **Introduction**

28

29 Weight of Evidence (WoE) is frequently cited as being necessary for a wide variety of decision-  
30 making needs due to the complexity of environmental data (Hall et al. 2017). Understanding  
31 environmental fate, hazard and exposure for any well studied chemical will normally reveal the  
32 presence of conflicting data and the presence of novel test systems and endpoints. A WoE  
33 analysis of such diverse data requires judgment by technical experts with a range of expertise  
34 and philosophical leanings. While standardization of testing procedures can help somewhat in  
35 this regard, as the knowledge of individual chemicals' properties expands or uses of a chemical  
36 changes, it is appropriate to tailor the assessment of the chemical to those aspects which possess  
37 the greatest likelihood of informing decision-making and not be overly narrow in the application  
38 of standard test procedures. New endpoints or understanding subtle aspects of how a chemical  
39 interacts with biota will inject novel information into the decision-making process. Well known  
40 examples of this phenomenon are the use and application of genomic techniques in human and  
41 environmental assessments and the emergence of endocrine disruption as an environmental  
42 concern. New information may cause scientists to reassess what they thought they already knew  
43 or provide new avenues to consider when assessing a chemical entity.

44 Another relatively new and major area (for environmental assessment) is the use of animal  
45 alternative assays, often referred to as New Approach Methodologies or NAMs, to address  
46 endpoints of regulatory concern. Lillicrap et al. (2016) reviewed the general state-of-the-science  
47 for various endpoints that require the use of fish for environmental assessment including  
48 bioaccumulation, acute and chronic toxicity and endocrine disruption. The acute fish toxicity  
49 (AFT) test (OECD 203, OECD 2019) is the most frequently used assay because it is a  
50 requirement in nearly all global regulatory schemes for the purposes of risk assessment and  
51 classification and labelling of chemicals (Burden et al. 2019). AFT tests account for the most fish  
52 used to satisfy regulatory testing requirements. While chronic (long-term) fish toxicity and  
53 bioaccumulation assays use more organisms per test, they are also used at a much lower  
54 frequency than AFT in the assessment of chemicals. Acute fish toxicity for effluent assessments  
55 in many countries utilize a large number of organisms and the employment of fish in research  
56 endeavors is a larger source of fish use as well (ECHA 2017). In our estimation, effluent testing

57 and research likely account for greater numbers than that where acute fish toxicity is a  
58 requirement for chemical registration (Norberg-King et al. 2018; ECHA 2017).

59 Bioethical concerns are a global driver for reducing the number of fish used in acute toxicity  
60 testing and replacing these tests with alternative *in vivo*, as well as *in vitro* and *in silico*  
61 approaches is a key objective. A wide range of regulatory, legal, and bioethical variability across  
62 the world makes the replacement of fish testing with NAMs extremely difficult. However,  
63 pressure on registrants of chemicals in commerce also continues to mount. In the absence of  
64 globally accepted recognition of replacement, reduction and refinement efforts, a reality is that  
65 acute fish toxicity tests may be required for registering a chemical in one region but not in  
66 another. Such dichotomies could be eliminated if evidence can be provided to convince  
67 scientists, and in particular regulatory authorities, that direct testing using juvenile fish (OECD  
68 203) is unnecessary to decide if fish are the most sensitive taxonomic group for determining  
69 hazard. Lammer et al. (2009) and Belanger et al. (2013) provided substantial evidence that  
70 toxicity of chemicals to fish embryos was equivalent to typical fish acute toxicity. This assertion  
71 was challenged by Scholz et al. (2016), resulting in a subsequent opinion rendered by the ECHA  
72 (European Chemicals Agency) (2017) to industry and interested scientists to produce a WoE  
73 approach to fully support the use of fish embryo toxicity data (and other alternative  
74 assays/approaches) in place of AFT data in European chemical registrations.

## 75 **Review of on-going work regarding AFT**

76 Efforts to address the execution, utility, and interpretation of alternatives to acute fish toxicity  
77 have been on-going for almost 30 years. Early efforts by Nagel (1994) and colleagues led to a  
78 formalized fish embryo test for effluent testing which was standardized as an ISO Guideline in  
79 the mid 2000's and subsequently adopted as an OECD Test Guideline in 2013 (OECD 236;  
80 OECD 2013). The fish embryo test relies on certain apical endpoints to accurately predict  
81 lethality including coagulation (death), lack of heartbeat, lack of somite development, and non-  
82 detachment of the tail. Lammer et al. (2009) and Belanger et al. (2013) compared the fish  
83 embryo toxicity (FET) to the AFT and found nearly a 1:1 concordance for a large number of  
84 chemicals across a wide range of modes of action (Figure 1). This work formed the basis of the  
85 assertion by some European chemical registrants to waive the AFT in lieu of the FET, for which

86 ECHA since clarified could be done but in a broader WoE context. As a complicating matter,  
87 recent simulation studies of FET versus AFT study designs clearly show that the OECD  
88 recommended minimum fish per concentration (n=7) compromises certainty in the LC50 and  
89 confidence limit estimates relative to the study design recommended for fish embryos (n=20 per  
90 concentration with positive and negative controls) (Carr et al. 2018). Recent efforts to improve  
91 the approach to AFT in the OECD 203 Test Guideline process includes utilization of clinical  
92 signs that predict mortality (i.e., moribundity) and shortening the test duration (Katsiadaki et al.  
93 2022).

94 Another alternative method is the implementation of the Threshold Approach (OECD 126,  
95 explained more fully below). The approach utilizes daphnid and algae toxicity tests to select the  
96 concentration at which to expose fish in a limit (threshold) test. Such an approach has been  
97 suggested as a means to reduce the use of fish by as much as 70% (Hutchinson et al. 2003).  
98 Although not explicitly expressed by Hutchinson et al. (2003), the Threshold Approach, can be  
99 considered a small WoE approach as it draws upon a variety of ecotoxicological insights to  
100 develop hazard conclusions.

101 Paparella et al. (2021) reviewed the known state of uncertainty and limitations of the acute fish  
102 toxicity test. The authors rightfully contend that the AFT has never been validated in the same  
103 sense that current OECD Test Guidelines require and as is outlined in OECD validation program  
104 documents under the Test Guidelines Programme (OECD 2005). The AFT, as currently  
105 practiced, has considerable drawbacks. Large variabilities in toxicity data for the same  
106 compound and species across laboratories are well documented in the literature (Belanger et al.  
107 2013) and yet, AFT results are normally viewed as the gold standard against which alternative  
108 assays are judged. Regulatory inertia, tradition, and a lack of familiarity with strengths and  
109 weaknesses provided by alternative assays all seem to play a significant role in the lack of  
110 regulatory acceptance to move beyond *in vivo* testing (Lillicrap et al. 2016). Many advantages  
111 are afforded by alternative assays including increased statistical power, improved interlaboratory  
112 calibration and validation for new, more robust assays, higher throughput, and improved  
113 mechanistic insights to the chemical's activity (Carr et al. 2019; Lillicrap et al. 2016, Paparella et  
114 al. 2021). Combining these additional tools in a WoE approach has the potential to significantly  
115 improve the environmental hazard and risk assessment of chemicals rather than relying on only

116 whether an organism simply lives or dies, which at the population level is too crude to ensure the  
117 protection of the environment. While acute toxicity may be useful for generating information on  
118 potency of various chemicals, it has lower relevance for environmental hazard assessment where  
119 longer term, lower-level exposures and prediction of long-term effects and environmental  
120 behaviors are more important. Yet, the AFT assay remains a mandatory requirement in most  
121 regulatory schemes.

122 AFT is but one of several *in vivo* fish assays that are used to inform environmental decisions of  
123 chemicals. QSARs, chronic (long-term) fish toxicity, bioaccumulation, endocrine disruption and  
124 field surveys all find a home in the comprehensive assessment of potential environmental  
125 perturbations and chemical exposures to fish. In 2012, the OECD brought together a group of  
126 scientists to consider the integration of all assays utilizing fish, with the goal of more efficiently  
127 using those assays to inform each other and reduce testing burdens resulting in the Fish  
128 Framework document (OECD 212). Fish embryo testing was foreseen as a future input into the  
129 fish assay toolkit (and was also identified as such as early as 2008 in guidance documents for  
130 REACH; ECHA 2008). Figure 2 displays one possible outcome of an integrated view on fish  
131 testing. As with the discussions above, this is also a form of WoE which is mentioned  
132 prominently throughout the Fish Framework document and its recommendations for future  
133 action. The OECD workshop identified that the following strategies could be implemented to  
134 reduce *in vivo* testing: limit tests, Threshold approach, step-down approach, screening  
135 methodologies that do not utilize animals, (such as (Q)SAR tools, *in vitro* assays, or read-across),  
136 and Fish embryo tests (FET). The FET is considered a replacement or refinement assay  
137 depending on the regulatory jurisdiction owing to the utilization of the non-exogenous feeding,  
138 embryonic stage of development (Strahle et al. 2012).

139 Clearly an integrative process is needed to holistically address the types of information and data  
140 available to inform the acute toxicity of chemicals to fish. The sources of information are  
141 diverse, vary in quantitative rigor, and chemical coverage (when considering for example,  
142 different types of alternative assays). In acknowledgement of this, Moe et al. (2020) and Lillicrap  
143 et al. (2020) initiated an effort to construct a quantitative model to integrate all sources of  
144 information using a Bayesian network approach. A preliminary version of the model is being  
145 evaluated as a proof of concept to inform development of a suitable structure that can be

146 amplified with increasing sophistication of modelling, chemical coverage, endpoints, and types  
147 of information that could be useful in the future. The goal is to provide a regulatory actionable  
148 decision-support system for WoE in acute fish toxicity. Formalized as a project under the CEFIC  
149 LRI Programme called SWiFT (**S**trengthening **W**eight of **E**vidence for Acute **F**ish **T**oxicity  
150 <https://www.niva.no/swift>), the Bayesian network model is a combination of tiered and  
151 probabilistic approaches. SWiFT intends to: 1) build a fully curated acute fish toxicity (AFT)  
152 database; 2) develop new (LoEs) for a Bayesian network (BN) model to predict acute fish  
153 toxicity; 3) develop and evaluate the BN model as a WoE framework to support AFT  
154 replacement; 4) develop a web interface for public access to the BN model; and, 5) produce  
155 guidance how to use the BN as a WoE tool.

### 156 **Conceptual description of AFT lines of evidence**

157 Predicting acute fish toxicity can draw from numerous LoEs and information. Typically, a  
158 complete understanding of a compound's physical and chemical properties is considered  
159 essential. Compound purity, solubility, molecular weight, sorptivity, pKa and other factors are  
160 ideally known and measured prior to testing (Schirmer et al. 2006). In addition, means to  
161 quantify in-test exposures under various water chemistry conditions is important as it is well  
162 known that exposure verification in *in vitro* and *in vivo* tests sheds important light on the  
163 comparability of measures between tests and across different compounds domains. That said, it  
164 is still uncommon to employ exposure verification, especially for difficult-to-test substances, and  
165 it is rare to summarize data for a compound where all possibly informative assays have  
166 equivalent levels of exposure verification (Belanger et al. 2013; Sobanska et al. 2018). This  
167 aspect alone has led researchers to employ varying levels of critical assessment of historical and  
168 current data when devising comparisons of alternative assays to the “gold standard” AFT (OECD  
169 236) (Paparella et al. 2021). In this aspect, one can see how WoE processes could be  
170 advantageous to sort out the most relevant and strongest central tendencies.

171 As mentioned previously, the FET test has been a major contributor for providing data to predict  
172 the AFT using an alternative approach (OECD 2013; Belanger et al. 2013). Similarly, an even  
173 more recent effort to establish the utility of a fish gill cell line to evaluate cytotoxicity of  
174 chemicals, and by inference predict acute fish toxicity, has been developed (ISO 2019; Fischer et

175 al. 2019; OECD 249, OECD 2021). Thus, three closely related assays (acute fish toxicity, fish  
176 gill cytotoxicity, and fish embryo toxicity) may be cross-referenced to inform acute fish toxicity  
177 of previously untested chemicals. Belanger et al. (2013), Tannenberger et al. (2013), Sobanska et  
178 al. (2018), and Fischer et al. (2019) each explored different aspects of the complexities involved  
179 through comparisons of outcomes amongst these assays. Belanger et al. (Figure 1) compared the  
180 toxicity of approximately 150 different compounds to FET and AFT (tests on the same  
181 compound) and found a near 1:1 concordance. However, variation in the effect data for the same  
182 chemical sometimes spanned orders of magnitude within either the AFT (more frequently)  
183 versus the FET. This is likely due to the heterogeneity particularly in the AFT data with respect  
184 to species choice (many species are recommended for use by OECD), test conditions, organism  
185 size, water source, and utilization of analytical (or not) confirmation. The comparisons were  
186 performed using orthogonal regression, a multivariate approach that allows for variation in both  
187 x and y dimensions (unlike standard linear regression where the independent x-variable has fixed  
188 values and the dependent y-variable is predicted with variation). Tannenberger et al. (2013)  
189 developed a different database where the AFT focused solely on fathead minnow, which  
190 constrains inter-species variability but simultaneously constrains AFT comparisons using species  
191 fully recommended and endorsed by OECD (OECD 2019). Figure 3A shows comparisons for  
192 cytotoxicity and *in vivo* toxicity for 37 compounds. Some, but not all, toxicity data in this Figure  
193 are also presented in that of Figure 1. Fischer et al. (2019), as part of the international round  
194 robin trial for the gill cytotoxicity assay, also summarized the comparative *in vitro* gill toxicity to  
195 the AFT (Fig. 3B). Again, as the gill cytotoxicity assay is relatively new, the breadth of coverage  
196 is less (albeit rapidly growing, cf. Natsch et al. 2018) and will partially intersect with that of the  
197 FET and AFT. Importantly, the FET and gill cytotoxicity assay have both undergone  
198 international validation and extensive ring trials to quantify their intra- and inter-laboratory  
199 variabilities as well as a demonstration of transferability to naïve laboratories and is now  
200 presently identified as OECD Test Guideline 249, “Fish Cell Line Acute Toxicity: The RTgill-  
201 W1 cell line assay”. (Busquet et al. 2014; Fischer et al. 2019; OECD 2021). Sobanska and  
202 colleagues presented a different view of the FET-AFT relationship that highlights some  
203 uncertainties in the newer assay (see Fig 4 and Sobanska et al. 2018). Quantification of chemical  
204 exposures identified a number of uncertainties which were subsequently resolved (Birke and  
205 Scholz 2019), but others remain, especially those with specific modes of action (see discussion



206 also below). As debated at the FET Workshop hosted by ECHA in 2017 (ECHA 2017), a  
207 mechanism is urgently needed to weigh results, predictions and ultimately derive predicted AFT  
208 when an *in vivo* test is not available to meet increasing expectations by society, regulators and  
209 scientists with regards to improved animal welfare.

210 An unrelated, but similarly complex module of a potential WoE scheme is that of the threshold  
211 approach (TA, OECD 126). The Threshold Approach, by itself, has also been posited as a WoE  
212 approach. In the Threshold Approach, acute toxicity tests are first performed on algae  
213 (unspecified species, but likely one of the small number of internationally accepted test species  
214 in OECD 201, OECD 2011) and *Daphnia* sp. (OECD 202, 2004). The more sensitive species of  
215 algae and *Daphnia* is identified by the lower of the two effect values (the concentration causing a  
216 50% effect - EC50). This concentration is then used as the test concentration in an acute toxicity  
217 limit test (i.e., 1 concentration at the threshold value of toxicity). If fish are not affected at this  
218 threshold concentration (i.e., 0% mortality), the assay confirms that fish are less sensitive and the  
219 assessor can proceed with confidence that the lower hazard value from the algae and daphnid  
220 tests will adequately inform the risk assessment. If fish are affected at the threshold  
221 concentration, then a full acute fish toxicity test is required. An easy extension to this thinking is  
222 to replace the AFT with that of the FET instead, thereby excluding the need for juvenile fish  
223 altogether. Rawlings et al. (2019) explored various permutations of algae-daphnid-AFT-FET  
224 testing to determine if the use of the FET or AFT impacted either the most sensitive hazard value  
225 or GHS classification. The authors conclude that there is no distinction (advantage) of using the  
226 AFT instead of the FET in the threshold approach and they can be considered equivalent. Such  
227 comparisons can also be used to assist testing prioritization and weightings applied to each assay  
228 for assessing overall acute fish toxicity. Rawlings and Belanger (personal communication,  
229 unpublished data) extended the threshold approach concepts to QSARs and found that while  
230 quantitative differences existed when using QSARs versus “real” data, the trends were  
231 remarkably similar albeit slightly higher uncertainty.

232 Scientists researching alternative methods consistently agree that identification of the mode of  
233 action (MOA) is an essential aspect of understanding toxicity relationships among chemicals and  
234 assays (Kienzler et al. 2017). Schirmer et al. (2006), Belanger et al. (2013), Tannenberger et al.  
235 (2013), Sobanska et al. (2018) and Fisher et al. (2019) use MOA assignments in parsing out

236 trends within the various databases used for comparing fish acute toxicity across compounds.  
237 This is particularly important as it is becoming well established that neurotoxicants represent a  
238 unique class of chemicals based on their MOA that are less well predicted by alternative assays  
239 (suggesting somewhat more frequently the FET and cytotoxicity tests, for example, are less  
240 sensitive to this particular mode of action) compared to the AFT. This is in conflict with the  
241 observation that neurotoxicants are also somewhat less toxic to fish overall than they are to  
242 invertebrates (Threshold Approach thinking, Connors et al. 2019). MOA is therefore critical in  
243 the development of WoE for predicting AFT. Kienzler et al. (2017) developed a MOA  
244 comparison for hundreds of compounds using several assignment schemes. These, like other  
245 databases, are built from different data sets, include different compounds, and were truly  
246 developed for different purposes or emphases. Subsequently, Kienzler et al. (2019) established a  
247 consensus approach to unify the MOA schemes with a single outcome based on physical-  
248 chemical information and multifaceted expert opinions, again forming a type of WoE.

249 Other factors may contribute to informing fish toxicity including collation of highly diverse fish  
250 metabolism and physiology information that can be useful to support MOA and understanding  
251 the potential for metabolic activation of unusual or specific compounds required to invoke  
252 toxicity to fish. While limited cases for such activation exist (allyl alcohol for example, see  
253 Kluver et al. 2014), these can explain outliers or trend busters for FET-AFT or cytotoxicity-AFT  
254 relationships (UBA 2020). Evolving neural toxicity assays, such as the quantification of the  
255 “touch-evoke” response or other behavioral endpoints using video tracking software, can more  
256 definitively determine the likelihood that a toxicant is or is not a neurotoxicant and thus lead to  
257 reliance on certain LoEs (e.g., the FET) versus others in predicting the AFT. The assignment of  
258 MOA will remain a significant area of research and will likely expand to include additional  
259 behavioral assessments such as swimming speed, endurance, and hyperactivity/hypoactivity.

## 260 **The Acute Fish Toxicity Bayesian Network Model as a Path to Resolution**

261 As indicated previously, Moe et al. (2020) and Lillicrap et al. (2020) provided the framework for  
262 developing a quantitative predictor of AFT using a Bayesian network (BN) model including  
263 multiple existing and proposed LoEs. BNs are graphical, probabilistic and potentially causal  
264 models, and have been used increasingly in hazard and risk assessment during the last decades

265 (Moe et al. 2021a). One of the strengths is the flexibility in model structure, which enables  
266 alignment of BN models with established frameworks such as adverse outcome pathways (Moe  
267 et al. 2021b) and the relative risk model (Landis 2021).

268 The BN model was developed using established WOE frameworks and guidance (e.g., Suter et  
269 al. 2017b, EFSA Scientific Committee et al. 2017) and is consistent with that described in this  
270 IEAM Special Series. The model gathers, weighs and integrates a wide range of LoEs that are  
271 used to develop a prediction of acute fish toxicity with a specified probability. A simplified  
272 graphical version of the preliminary model is shown in Figure 5. A database was built using all  
273 possible chemicals for which data on an alternative biological assay, relevant to AFT, are  
274 available. If only AFT and QSAR data were available for a compound, it is not considered in this  
275 database until an alternative assay is performed. The goal is to provide predictive support for  
276 acute fish toxicity using animal alternative toxicity tests. An expectation of the BN approach was  
277 that inclusion of data from more LoEs would result in a more accurate AFT prediction (that is, its  
278 central tendency being closer to the measured AFT). Model accuracy was defined by correctly  
279 predicting the toxicity interval of the AFT data using multiple LoEs. The accuracy rate of the BN  
280 model prediction was in the range of 69-80% when using available data for all LoEs as far as  
281 possible. A diverse array of chemicals and MOAs was used to test model predictions to assess if  
282 the model could be broadly applicable (Moe et al 2020). The strictest quality criterion resulted in  
283 a set of 20 chemicals which included various industrial compounds, surfactants, pesticides, and  
284 pharmaceuticals (see Table 2 of Lillicrap et al. 2020). For this subset, the BN predicted the  
285 correct toxicity interval for 80% of the chemicals evaluated. For the remaining 20% of the  
286 chemicals, daphnid or algae data were always more sensitive than fish, which means that  
287 daphnid or algae data would have driven any subsequent environmental hazard assessment or  
288 GHS classification. The evaluation also confirmed that the use of FET data to replace AFT data  
289 was justified in that GHS classification or toxicity interval predictions were the same.

290 Table 1 provides a summary of potential LoEs and their present likely strengths to support a  
291 WoE determination of AFT. Fish acute toxicity QSARs vary widely in the breadth of underlying  
292 data used in their development and the predictive chemical attribute also varies, but is most  
293 commonly based on log Kow. Through on-going efforts, such as those of EU JRC QSAR  
294 validation requirements for REACH (OECD 2004; EU JRC 2014), the aim is to provide a

295 baseline from which valid and accepted QSAR models can be derived and used for the BN. FET  
296 and fish gill cytotoxicity are considered strong, direct evidence in support of AFT based on  
297 earlier reviews of these assays. The Threshold Approach (TA) , as described earlier, is  
298 considered supportive and indirect evidence of AFT. The outcomes of the TA is frequently  
299 more conservative (lower LC/EC50) than for the full acute fish toxicity test. When used in  
300 combination with other LoEs the TA can further support AFT conclusions. MOA assignment is  
301 particularly challenging as several accepted classification systems exist. For the further  
302 development of BN, the integrated consensus approach given in EnviroTox  
303 (<http://www.envirotoxdatabase.org/>) is utilized (see Kienzler et al 2019). MOA can be useful to  
304 identify compounds that are likely more toxic to fish (e.g., selected neurotoxicants) or other  
305 organisms (e.g., herbicides being more toxic to algae). This can feed into the threshold approach  
306 for supporting indirect LoEs and allows investigators to “make sense” of either variably toxic  
307 compounds or the likelihood that a single individual assay is correct. Newer LoEs such as  
308 gathering knowledge of biochemical and metabolic pathways or behavioral/physiological assays  
309 can be used to inform the potential of metabolism (often reducing internal toxic burdens for a  
310 compound), metabolic activation (making a compound more toxic potentially), and whether  
311 these fit into existing MOA frameworks.

312 Here, we have explored more systematically the importance of the number of LoEs in data-rich  
313 vs. data-poor scenarios using two example substances (Figure 6): Triclosan (high toxicity to fish)  
314 and tetradecyl sulphate (medium toxicity). For each LoE, the BN model uses either evidence  
315 (data) for the given substance or a prior probability distribution based on all training data,  
316 depending on the scenario. In the example with triclosan, all scenarios with evidence for four or  
317 three LOEs give the correct model prediction as indicated by high quality AFT results. When  
318 evidence for only two LoEs are used, the toxicity level is underestimated in 2 out of 6 cases.  
319 When evidence for only one LoE is used, toxicity is even more strongly underestimated in 2 out  
320 of 4 cases. The BN predicts the correct interval for all scenarios where embryo data are used. In  
321 the case of tetradecyl sulfate, the prediction accuracy (correct interval have highest probability)  
322 increases with the number of LoEs with evidence. Compared to triclosan, the predictions for  
323 tetradecyl sulfate tend to have lower precision (the peak probabilities don't get as high) and a bit  
324 lower accuracy (only 8 out of 15 have correct interval, compared to 11 out of 15 for triclosan).

325 This is likely related to the fact that more toxicity values for algae (n=4) and Daphnia (n=6) are  
326 available for triclosan.

327 To generalize from these two examples, inclusion of evidence from a larger number of LoEs in  
328 this BN model seems to improve both accuracy (more cases of correctly predicted toxicity  
329 interval) and precision (higher probability of the predicted most probable interval). We also  
330 expect that inclusion of more replicate values (repetitive tests on the same endpoints) within an  
331 LoE would give more precise AFT predictions (that is, narrower probability distributions). This  
332 aspect will be further analyzed in the current SWiFT project. Actual AFT data is used within the  
333 model evaluation. Of course, this assumes that the AFT data is precise and accurate, therefore,  
334 an effort to assess reliability of toxicity tests and curate detailed results is also part of the effort.  
335 It is important to recognize that the BN model utilizes all data to understand variance in  
336 measured endpoints. This is an improvement over the common practice of using geometric mean  
337 values to summarize multiple measurements for the same taxon (Stephan et al. 1985; ECHA  
338 2008).

339 Figure 5 displays how LoEs are combined in the BN model. Within each LoE (pathway), toxicity  
340 to fish is predicted from one or more nodes. The toxicity nodes are arbitrarily discretized into 5  
341 toxicity levels, based on intervals of effect concentration (LC50): very low (>100 mg/L), low (5–  
342 100 mg/L), medium (0.5–5 mg/L), high (0.5–0.01 mg/L), and very high (<0.01 mg/L). These  
343 toxicity levels are relevant since aquatic toxicity not only used as environmental risk assessment  
344 inputs for PNEC (Predicted No Effect Concentration for ecosystems) determination, but also for  
345 classification and labelling under the Globally Harmonized System. For all parent nodes (with no  
346 incoming arrows), prior probability distributions were calculated based on all substances in the  
347 dataset. Each child node has a conditional probability table (CPT) that is used for calculating the  
348 posterior probability distribution dependent on the probability distribution of the parent node,  
349 when the model is run for a given chemical. The CPTs are constructed both from expert  
350 judgement (e.g., assignment of chemical categories) and from empirical relationships (e.g.,  
351 frequency distributions of toxicity levels in different chemical categories).

352 Table 1 (inputs for the BN) can be thought of in a tiered approach quite easily. Direct LoE  
353 provide the best evidence for AFT, assuming a high-quality assay is performed. The more LoE

354 employed, the higher the tier. Outputs are provided along the way as a probability for a particular  
355 outcome. This is a similar process to the historical tiering of aquatic toxicity data given in Cairns  
356 and Dickson (1978) and described in Menzie et al. (2021). The more LoEs that are implemented  
357 and the more evidence within each LoE that is gathered, both the accuracy and precision of the  
358 prediction should ideally improve. As with conventional risk assessment paradigms, the level of  
359 uncertainty that can be tolerated in a given environmental decision is related to the magnitude of  
360 the difference between hazard and exposure concentrations and the precision required for  
361 acceptance.

362 The AFT BN model is a quantitative approach to WoE (while many others are qualitative, see  
363 Suter et al. 2017). The BN can conceivably be mapped to a WoE framework. The weighting in a  
364 traditional WoE could be implemented as probability distributions in a BN, both within  
365 individual LoEs and for integrating the LoEs (Moe et al. 2021). The assignment of conditional  
366 probabilities to different variables within in a LoE in the BN corresponds to setting weights to  
367 pieces of evidence in a WoE (Moe et al. 2021). Assigning a lower weight to a piece of evidence  
368 can be obtained by setting wide probability distributions (representing high uncertainty or  
369 variability) in the relationship from this node to its child node. Within a line of evidence the  
370 calculation of posterior probabilities for the last child node of the line then accumulates the  
371 weights given to all parent nodes in this line. It follows that when using the BN to predict the  
372 toxicity to juvenile fish from all LoE for a chemical of concern, the weighting of the total  
373 evidence for each hypothesis in a WoE (Suter et al., 2017a) can correspond to calculating the  
374 posterior probability of each toxicity level (“very low”, “low” etc.) in the BN.

### 375 **What a successful WoE for AFT Looks Like**

376 Regulatory acceptance of animal alternative approaches is challenging. There are many reasons  
377 for this including the structure of regulations which vary across the globe, the dependence on  
378 vertebrate data in historical assessments which could be “re-visited” if reliance on alternatives  
379 take hold, the uneven levels of expertise in the scientific and regulatory communities, and fast-  
380 paced evolving regulatory changes which “move, and often raise, the bar”. Due to these and  
381 many other factors, a successful WoE for AFT would possess the following attributes:

- 382 1. The approach would have global applicability;

- 383 2. Outcomes would be met with regulatory acceptance and not require significant changes  
384 to legislations or federal law;
- 385 3. Would be relevant equally for all fish species whose use varies across regulatory  
386 jurisdictions (e.g., Europe employs zebrafish most frequently, the US uses fathead  
387 minnow, Canada and the UK rely heavily upon rainbow trout, Japan uses Medaka  
388 preferentially and movement exists for China to rely upon the Chinese rare minnow).
- 389 4. The model leaves open for expansion of knowledge regarding chemical domain, new  
390 LoEs, and underlying input data;
- 391 5. Would include a renewed recognition that the “gold standard” AFT assay, is not  
392 necessarily more precise and accurate, thus, improving the transparency and curation of  
393 accepted highest quality AFT data is critical;
- 394 6. Would improve the environmental hazard and risk assessment of chemicals;
- 395 7. Be publicly available, but controlled, to ensure transparent application.

396 Beyond AFT, other areas of fish hazard assessment could benefit from a similar approach (and  
397 some are already in progress). Bioaccumulation assessment is identified in ECHA (2008, 2017)  
398 as being a WoE process and is discussed in more detail within this special issue by Arnot and  
399 colleagues (Arnot et al. 2021, in this Special Series) and previously by Lillicrap et al. (2016).  
400 Endocrine disruption and long-term fish toxicity were identified in OECD (2012) as also  
401 requiring WoE development. Bioaccumulation, endocrine disruption (also discussed by Mihaich  
402 and Burgoon et al. (2021) (in this Special Series) and long-term fish toxicity all involve  
403 assessment of complex endpoints that require deep expertise. A transparent, accepted WoE  
404 process for all would benefit the regulatory environment in addition to the early development of  
405 new chemical entities. Furthermore, animal alternative assays will play a pivotal role in future  
406 WoE approaches as already identified by Volz et al. 2011 and Villeneuve et al. 2014 for long-  
407 term fish toxicity, Nichols et al. (2018) for bioaccumulation and Scholz et al. (2014) for potential  
408 endocrine disruption. In all of these, the guiding principles for WoE will be the same but a much  
409 wider net of lines of evidence will likely be needed. Whether Bayesian approaches would fulfill  
410 some of these needs is yet to be answered.

411 Many remaining challenges exist, including how to deal with novel chemicals which may not  
412 have much context for their use, MOA and properties. That said, the surface has been scratched

413 with respect to existing compounds whose assessments are part of the historical record, but lack  
414 animal alternative data. Compounds with difficult solubility, sorptivity, analytical verification  
415 and toxic properties will also remain challenging. Importantly, regulators and scientists will need  
416 to possess courage to make decisions when data are less than perfect (which will always be the  
417 situation) and develop new regulatory inertia in the face of a lack of precedents. Finally, since  
418 environmental risk assessment frameworks and chemical legislations are principally employed  
419 for the protection of the environment, hazard assessment strategies should be moving away from  
420 relying on acute fish toxicity for classification purposes and focus more on mechanistic/sublethal  
421 endpoints to identify chemicals of concern.

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644

645 Table 1. Strength of attributes presently incorporated or being considered in the BN for the AFT  
 646

Attribute	Relevant Test Guideline(s)	Measured endpoint	Strength (potential weight or inference towards AFT)	Status in Moe et al. 2020 BN	Utility for Tiered Weight of Evidence
Physical-chemical properties	Numerous, compound dependent	Solubility, log Kow, pKa, molecular weight	Strong, influence on MOA	Included	Essential
Mode of Action	Scientific judgement	Assignment of mode	Strong	Included, see Kienzler et al. (2017) and <a href="http://www.EnviroToxdatabase.org">www.EnviroToxdatabase.org</a>	Important
Measured Acute Fish Toxicity	OECD 203	96 h LC50	Direct evidence	Used as context for similarly acting (MoA) chemicals (in the line "Chemical category")	Direct evidence, may still need to weigh conflicting data (variability in test results; use of various species)
Fish QSAR	Various accepted platforms (ECOSAR, Danish QSAR, OECD Toolbox)	96 h LC50	Strong when QSARs are validated and chemical is within domain of applicability	Included in the line "Chemical properties"	Indirect evidence
Measured Acute Fish Embryo Toxicity	OECD 236	96 h LC50	Strong	Included in the line "Embryo"	Direct evidence
Measured Acute <i>Daphnia</i> Toxicity	OECD 202	48 h EC50	Must be used in combination with other attributes; potential applicability in Threshold Approach	Included in the line "Other taxa"	Indirect evidence when used in combination with algal inhibition
<i>Daphnia</i> QSAR	ECOSAR, OECD	48 EC50	Strong when QSARs are	Candidate	Indirect evidence

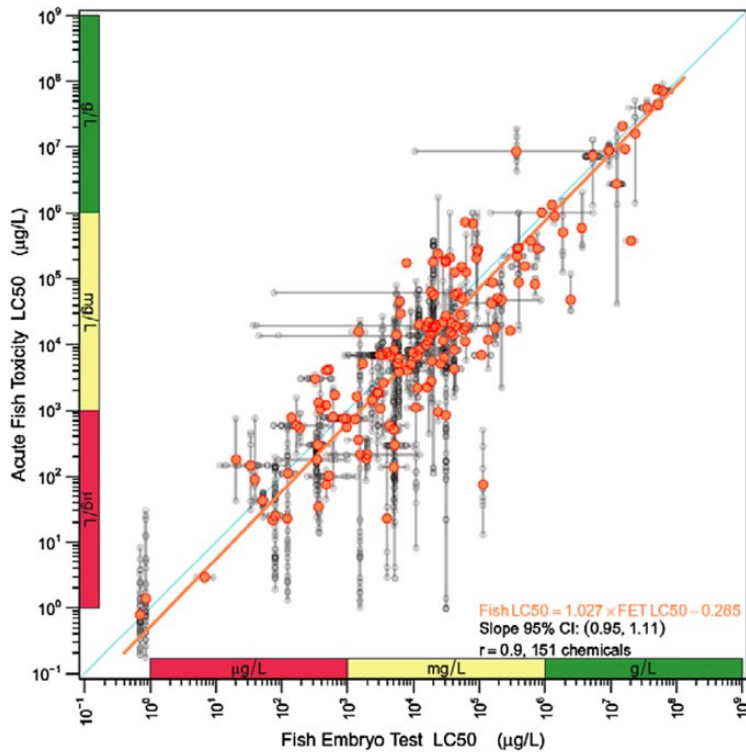
	QSAR Toolbox		validated and chemical is within domain of applicability; must be linked by other means to fish toxicity (TA)		when used in combination with algal inhibition
Measured Algal acute inhibition	OECD 201	72 h ErC50 96 h EC50	Moderate; numerous species and complexities of regulatory endpoints <i>sensu</i> Brill et al. 2021; useful/necessary within TA	Included in the line "Other taxa"	Indirect evidence when used in combination with algal inhibition
Algae QSAR	ECOSAR, OECD QSAR Toolbox	72 h EC50 96 h EC50	Moderate; few broadly available QSARs	Candidate	Indirect evidence when used in combination with algal inhibition
Fish gill cytotoxicity	OECD 249 ISO 21115	24 h EC50	Moderate; growing database, prospects are very good	To be included	Inferential, depends on MoA
Mode of Action assignment	EnviroTox	N/A	Complexity across MoA schemes (Kienzler et al. 2017); Important to identify specifically-acting chemicals	Included in the line "Other taxa", to be expanded	Supportive
Neurotoxicant behavior assay	In development	Neural impact, Yes/No	In progress (Touch evocation assay); important means to identify this class	In exploration	Supportive



Fish metabolism	Few standardized assays; OECD 319a,b; Km QSARs	Various biochemical and physiological measurements	In development; much is known but not fully catalogued (Braunbeck et al. 2019, Loerracher and Braunbeck 2021)	In exploration	Supportive
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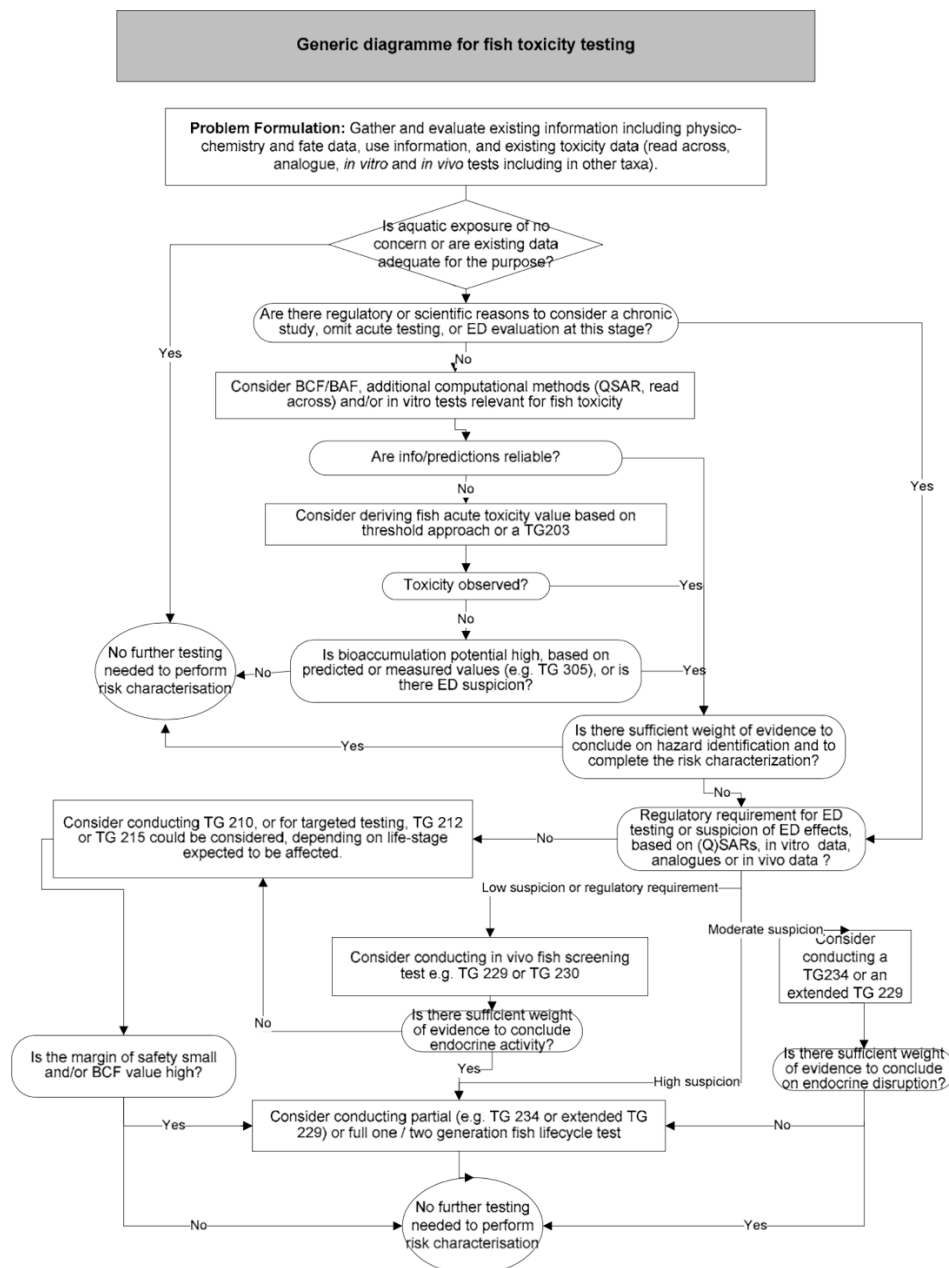
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Figure 1. Comparison of fish embryo test (FET) versus acute fish toxicity (AFT) for 151 compounds from Belanger et al. (2013).

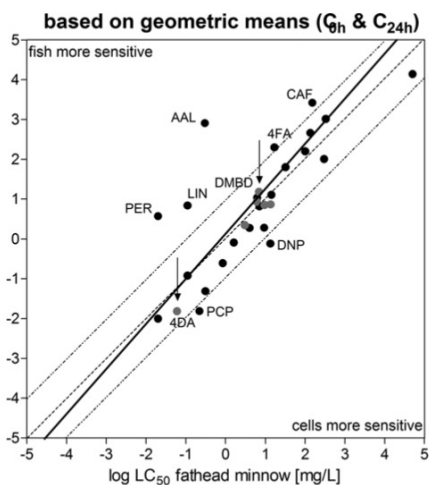


**Figure 7.1:** A generic testing strategy illustrating general principles of how to optimise fish toxicity testing needs. This strategy makes use of as much prior information as possible in order to determine the need for fish testing. Regulatory requirements in particular jurisdictions may require a more complex or testing-rich assessment. Risk characterization and other chemical assessments may include, for example, a PBT assessment, hazard classification, and / or endocrine assessment.

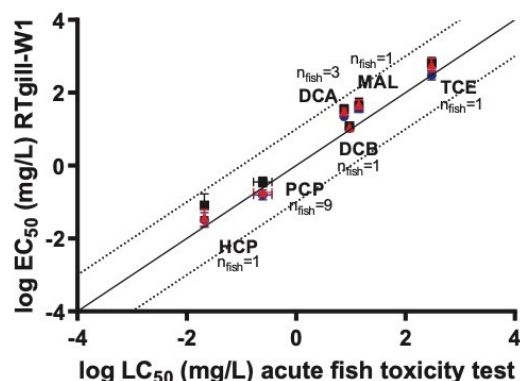
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656 Figure 2. An integrated approach to informing the environmental hazards of chemicals to fish  
657 (Reproduced from the OECD Fish Framework (OECD 2012)).

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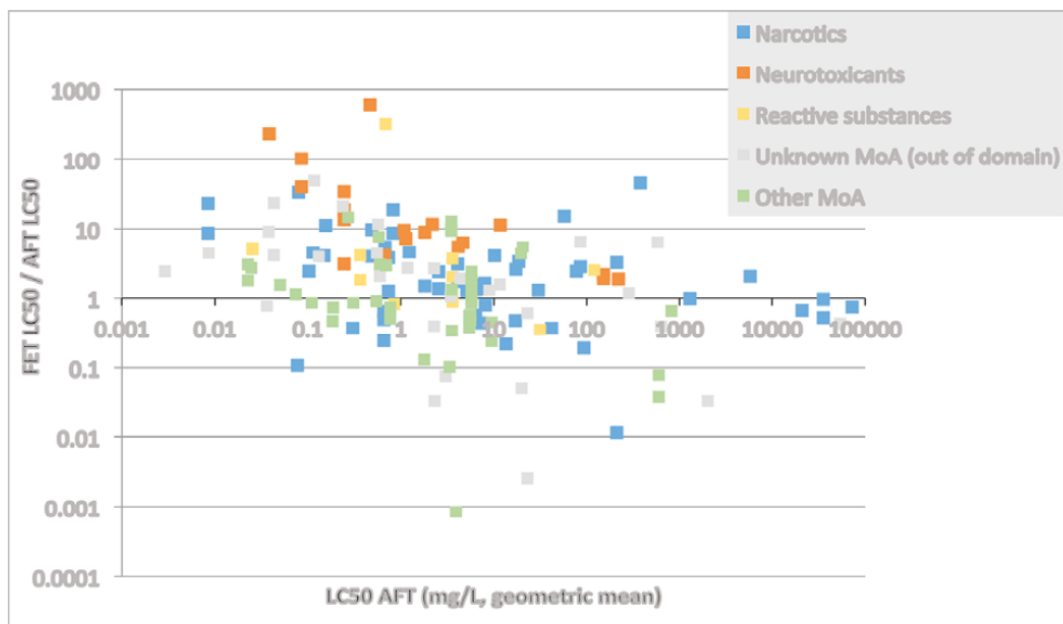


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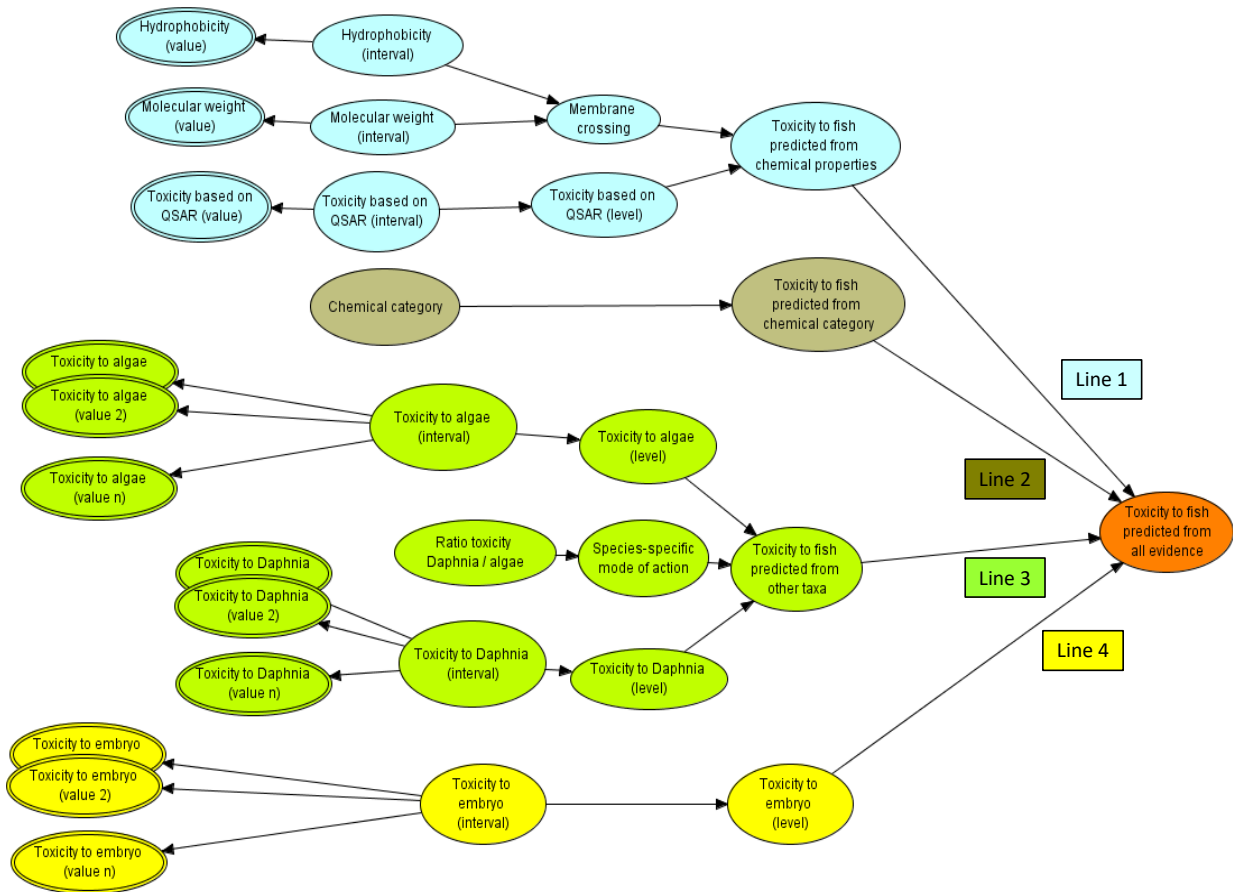


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Figure 3. (A) Linear correlations between acute fathead minnow toxicity and fish cell line EC50 values for metabolic activity as given in Tannenberger et al. (2013). (B) Correspondence of round-robin study-derived average log EC50 values obtained for the 3 different cell viability measurements per test chemical with average log LC50 values from fish acute toxicity testing as given in Fischer et al. (2019). Correspondence of the cell line derived data with *in vivo* data obtained from the US EPA fathead minnow database (see Russom et al. 1997 and Tanneberger et al. 2013).

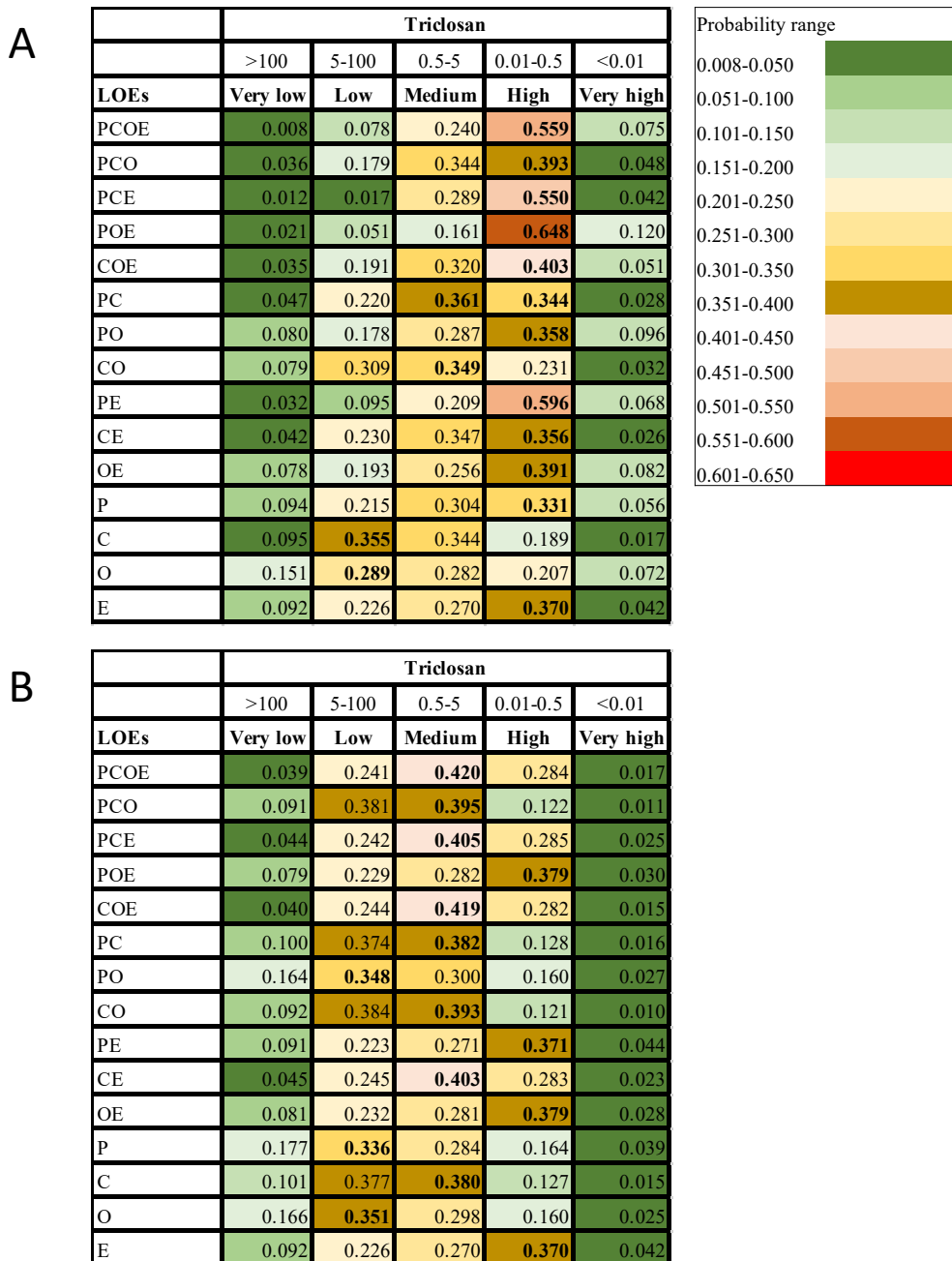


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 669 Figure 4. Differences between fish embryo toxicity (FET) and acute fish toxicity (AFT) effect  
 670 concentrations as described by Sobanska et al. (2018). The ratio between FET and AFT (y-axis)  
 671 over the range of median lethal concentrations (LC50s) in the acute fish toxicity (x-axis).  
 672 Different modes of action are indicated in different legend colors.  
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Figure 5. Conceptual version of the Bayesian network model to predict Acute Fish Toxicity from four lines of evidence (after Moe et al. 2020). Nodes with double outlines represent input values (observed toxicity values) in continuous scale; the model currently accepts up to 10 replicate observations. All other nodes have discrete states, such as concentration intervals.



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Figure 6. Comparison of BN model predictions of fish toxicity under data-rich and data-poor scenarios for two example substances: (a) Triclosan and (b) Tetradecyl sulphate. The 15 scenarios use available data for one, two three or all four of the lines of evidence (LOEs): P = physical/chemical properties, C = chemical category, O = other taxa (algae and *Daphnia*), E = fish embryo. For each scenario, the table shows the predicted probability distribution across the five toxicity levels, defined by intervals of LC50 values (in mg/L). The outlined column indicates the observed toxicity interval, while bold numbers highlight the interval with the highest predicted probability. Colors indicate broad ranges of probability of occurrence ranging from dark green (low) to red (high).