

23 **Abstract**

24 Wastewater from certain industrial processes can be primary emission sources of per-
25 and polyfluoroalkyl substances (PFAS) and fluorinated alternatives like chlorinated
26 polyfluorinated ether sulfonates (Cl-PFESA). Two such industrial processes are
27 electroplating and textile printing and dyeing (PD). This study focused on the fate of
28 Cl-PFESA in wastewater from these two industrial processes, in comparison to other
29 PFAS, as they went through different wastewater treatment plants located in southeast
30 China. The total target PFAS concentrations were 520 ± 30 ng/L and 4200 ± 270 ng/L at
31 the effluents of the PD WWTP and electroplating WWTP, respectively. Specifically,
32 6:2 Cl-PFESA (18%) and 8:2 Cl-PFESA (0.7%) were abundant in electroplating-
33 wastewater. Cl-PFESA were also detected in PD wastewater but at trace concentrations
34 and were likely present due to diffuse emissions. The dissolved-phase Cl-PFESA and
35 PFAS mass flow through the WWTPs were fairly constant throughout both facilities.
36 The majority of Cl-PFESA was captured by sludge sedimentation. However, there were
37 individual treatment processes that could cause the wastewater concentrations to
38 fluctuate, and also could lead to relative enrichment of specific Cl-PFESA as indicated
39 by the 6:2/8:2 Cl-PFESA ratios. Cl-PFESA and perfluoroalkyl sulfonic acids were more
40 influenced by the investigated treatment processes than perfluorocarboxylic acids.

41 **Keywords:** PFAS; industrial wastewater; water pollution; source apportion

42 **Synopsis:** Fate behavior of chlorinated polyfluorinated ether sulfonates and other PFAS
43 were characterized in different industrial wastewater treatment plants

44

45 **Introduction**

46 Over 4500 per-and polyfluoroalkyl substances (PFAS) exist, and many have been used
47 in various industrial applications and materials found in numerous customer products
48 including papers, textiles, plastics and glass, often to obtain nonstick, water-repellence
49 and low-friction properties^{1,2}. Perfluoroalkyl sulfonic acids (PFASs) and
50 perfluorocarboxylic acids (PFCAs) are the two main groups of PFAS; however, recently
51 due to both increased regulatory action and market forces, new types of fluorinated
52 alternatives are emerging on the market, such as chlorinated polyfluorinated ether
53 sulfonate acids (Cl-PFESAs). Numerous studies have demonstrated that PFAS are wide
54 spread in natural aquatic systems, including rivers^{3,4}, lakes^{5,6}, groundwater^{7,8} and sea
55 water^{9,10}. Due to their impact on aquatic environments as well as drinking water
56 resources, water contamination by PFAS has been an issue of growing public concern¹¹⁻
57 ¹³.

58 PFAS have been intensively applied in many local industrial activities, where they can
59 become long-term PFAS hotspots for local environments. Studies of PFAS entering
60 wastewater treatment plants (WWTPs)¹⁴⁻¹⁸ have indicated that industrial wastewater
61 can be a substantial source of emissions, particularly those from the electroplating¹⁹,
62 textile manufacturing²⁰ and chemical production industries^{21,22}. During the last decade
63 PFAS production in China has undergone rapid growth since the restriction and
64 reduction of perfluorooctyl sulfonate (PFOS) and/or perfluorooctanoic acid (PFOA)
65 manufacturing in the U.S. and Europe. In China, PFOS has been in high demand for
66 the chrome plating industry, where PFOS and its salts are applied as chrome mist

67 suppressants to prevent the formation of chrome-containing mist²³. For almost 40 years
68 monochlorinated polyfluorinated ether sulfonate (Cl-PFESA), particularly 6:2 Cl-
69 PFESA often called F53B, have been applied as PFOS alternatives in the metal plating
70 industries due to their simpler chemical production procedures as well as lower
71 production costs^{23, 24}. A recent modeling study estimated that 10–14 tonnes of F53B
72 have been released annually to the environment in China during 2006–2015²⁴. Another
73 active PFAS hotspot is textile manufacturing. Side-chain fluorinated polymers or
74 perfluorooctane sulfonyl fluoride (POSF)-based derivatives were commonly used for
75 surface treatment of textiles in order to obtain water-proof and oil-resistant properties.
76 An increasing trend in China was to use shorter-chained homologues as replacement
77 chemicals²⁵. For instance, perfluorohexane sulfonyl fluoride (PHxSF) derivatives have
78 been increasingly produced as alternative surface treatment chemicals; these can
79 notably degrade into perfluorohexane sulfonate (PFHxS)^{25, 26}. Due to the changing
80 composition of PFAS alternatives as surface treatment chemicals, their fate and that of
81 their degradation products in wastewater still requires better characterization.

82 Different industrial wastewaters often have special emission inventories covering both
83 legacy and alternative PFAS, and therefore might serve as an identifier for PFAS
84 discharge from different sources. PFAS concentrations from these different sources
85 could vary temporally and spatially during individual wastewater treatment, depending
86 on the physiochemical properties of different PFAS as well as their sensitivity to
87 different treatment techniques^{27, 28}. Though industrial wastewater treatment plants
88 directly receive PFAS containing wastewaters, water treatment facilities are rarely

89 designed for PFAS removal²⁹. WWTP effluents and sludge are considered active
90 sources for the environmental release of both legacy and alternative PFAS³⁰. In China
91 Cl-PFESA like F53B represents one of the most important country-specific PFOS
92 alternatives. There is increasing evidence of the emergence of 6:2 Cl-PFESA and 8:2
93 Cl-PFESA in both wastewaters^{15, 31, 32} and fresh water environments in China^{12, 33}.
94 However, the knowledge on diverse industrial uses, in particular, for small-scale
95 manufacturers, and local emission hotspots of Cl-PFESA are still lacking. Hence, there
96 is growing interests to identify different sources of Cl-PFESA and to understand their
97 fate in industrial wastewater streams in comparison to that of legacy PFAS. The main
98 objectives of this study are to characterize the fate and transport of 6:2 Cl-PFESA and
99 8:2 Cl-PFESA from the two different in-situ industrial wastewater treatment plants
100 receiving wastewaters from electroplating and textile printing and dyeing manufacturing,
101 respectively, and comparing them with legacy PFAS.

102

103 **Materials and methods**

104 **Chemicals and Reagents**

105 In total eighteen PFAS were included as target compounds. Six isotope-labeled PFAS
106 were employed as internal standards. All the standards were obtained from Wellington
107 Laboratories (Ontario, Canada), with purities greater than 98%. Detailed information
108 regarding the target analytes are summarized in Table S1. Millipore water was produced
109 by a Milli-Q system from Sartorius (Göttingen, Germany). LC-MS grade solvent (i.e.,
110 methanol and ammonium hydroxide; v/v, 25%) were purchased from Merck (Darmstadt,
111 Germany). Glass fiber filters (GFFs, 47 mm diameter, 0.7 μm aperture) were obtained

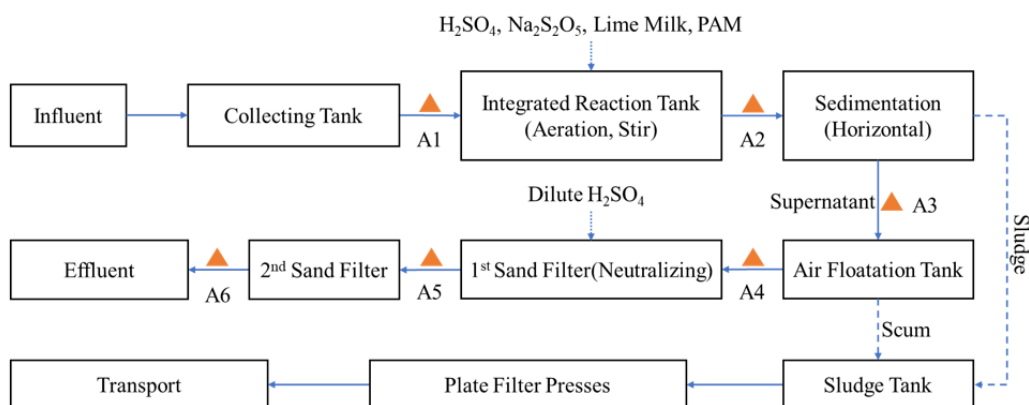
112 from Whatman (Maidstone, UK). Oasis weak anion exchange cartridges (WAX, 150
113 mg, 6 mL, 30 μm) were purchased from Waters (Milford, USA) and Envi-Carb
114 cartridge (3 mL, 250 mg) was purchased from Supelco (Bellefonte, USA).

115 **Sample Collection**

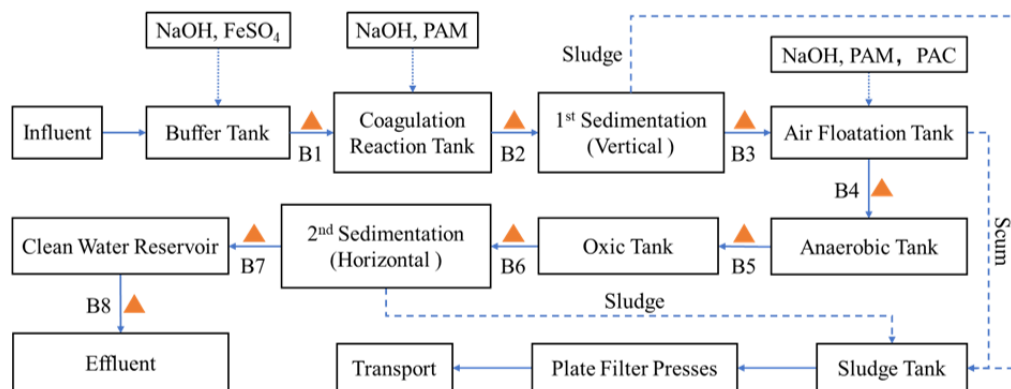
116 Industrial wastewater from an electroplating wastewater treatment plant (E-WWTP)
117 and a textile printing and dyeing wastewater treatment plant (PD-WWTP) were
118 separately collected during one day in November 2019. Both WWTPs are located in
119 the Pearl River basin in southeast China, where numerous small-scale industries are
120 located. The E-WWTP is treating wastewater collected from surrounding chrome
121 plating factories, and mainly implements chemical precipitation to treat the industrial
122 wastewater. PD-WWTP is exclusively collecting wastewater generated during textile
123 printing and dyeing processes in several local factories. The main treatment processes
124 of PD-WWTP is chemical precipitation (via flocculation), anaerobic digestion and then
125 aerobic digestion. The flocculent used in both facilities is polyacrylamide (PAM)^{34, 35}.
126 The detailed description of the two WWTPs are shown in Figure 1 and Table S2a.
127 E-WWTP and PD-WWTP are treating a fixed amount (i.e. 1000 m^3/d and 330 m^3/d ,
128 respectively) of the collected industrial wastewater, which goes once through the entire
129 treatment procedures daily. The wastewater sampling followed the wastewater stream
130 by considering hydraulic retention time during individual treatment step, and therefore
131 allowed for tracking the fate of the target PFAS along the wastewater stream as well as
132 for substantial flow analysis. Though this sampling regime does not give time-
133 integrated samples, it does prevent biases caused by temporal fluctuations when not

134 following the hydraulic retention time (see Tables S2b and S2c) through the WWTP³⁶.
 135 Treated wastewater was collected in a 1L polypropylene (PP) bottle from the effluent
 136 of different treatment units in both WWTPs. All sampling bottles were washed with
 137 methanol and rinsed three times with wastewater sample before collection, and sealed
 138 with sealing film immediately after sampling. Prior to analysis the samples were stored
 139 at 4°C room in the dark. The sludge that accumulated during the treatment processes
 140 were sampled (about 250 g per wet sludge sample) before the dehydration treatment.
 141 The wet sludge sample was immediately packed with aluminum foil and sealed in
 142 polypropylene bags. Prior to analysis, the sludge samples were preserved at -20°C.

a. Electroplating wastewater treatment plant.



b. Printing and dyeing wastewater treatment plant.



143

144 Figure 1. Water treatment processes of the two in-situ industrial wastewater treatment plants.

145 Panel (a) the E-WWTP and Panel (b) the PD-WWTP. The yellow triangle represents the
146 sampling locations. The dashed lines represent the removal of sludge from various wastewater
147 treatment steps.

148 **Sample pretreatment**

149 **Filtration.** 500 mL of wastewater samples were filtered by glass fiber filters (0.7 μm ,
150 Whatman, Maidstone, UK), then sealed in PP bottles and stored at 4 °C. All filters were
151 baked at 450 °C for 12 hours before use. Given that PFAS sorption on glass fiber filters
152 might underestimate PFAS concentrations^{37, 38}, after the filtration of water sample, the
153 filters were washed with about 15 mL methanol to extract PFAS residues on the glass
154 filters as it was recommended in a previous study³⁷, and then the methanol solution was
155 collected and combined later added to the treated water samples extracted in methanol
156 described below. The sludge samples were pretreated using the dispersive solid phase
157 extraction (DSPE) method. The detailed treatment protocols were based on a previous
158 study.³⁹

159 **Extraction.** The aqueous samples were extracted by solid phase extraction (SPE) using
160 Oasis WAX cartridges. Before water loading, samples were spiked with 5 ng ISs (50
161 μL , 100 $\text{pg}/\mu\text{L}$). The WAX cartridges were activated with 10 mL acetone, 10 mL
162 methanol and 10 mL 0.25 % ammonium hydroxide in methanol before use. The loading
163 speed was adjusted at a flow rate of approximately 2 mL/min.

164 After water loading, the cartridges were cleaned with 5 mL Millipore water and dried
165 with a vacuum pump. Then, dried cartridges were eluted with 15 mL 2.5% ammonium
166 hydroxide in methanol. The eluents were kept in preconditioned PP centrifuge tubes

167 (50 mL, Biosharp). After purification by Envi-Carb cartridges, the eluents were
168 concentrated to 190 μ L under a gentle stream of nitrogen. The Envi-Carb cartridges
169 were cleaned with 15 mL methanol. Before instrumental analysis, 1 ng (10 μ L, 100
170 pg/ μ L) injection standards ($[^{13}\text{C}_2]$ -PFOA) were added in each sample.

171

172 **Instrumental analysis**

173 Samples were analyzed by an ultrahigh performance liquid chromatography tandem
174 mass spectrometry (UHPLC-MS/MS) system using a 1290 Infinity II UHPLC (Agilent
175 Technologies) with a 6470 Triple Quad MS/MS (Agilent Technologies). The analytical
176 column used was a ZORBAX Eclipse Plus C18 column (2.1 \times 100 mm, 1.8-micron,
177 Agilent Technologies) with a UHPLC Guard for C18 (2.1 \times 50 mm, 1.8-micron, Agilent
178 Technologies). The column temperature was kept at 40 $^{\circ}$ C. The mobile phase consisted
179 of A: 10 mmol \cdot L $^{-1}$ ammonium acetate in water and B: 10 mmol \cdot L $^{-1}$ ammonium acetate
180 in methanol. The gradient profile was achieved at a flow rate of 0.30 mL/min and
181 initiated with an equilibration of 90% A, which was decreased to 35% A for 5.5 min
182 and held for 0.5 min, then decreased to 10% A for 2.5 min and held for 3.1 min, and
183 then increased to 90% A for 0.1 min. Finally, 90% A was held for 0.8 min. The detailed
184 instrument parameters of the UHPLC-MS/MS were presented in Table S3.

185

186 **Quality assurance and quality control**

187 To prevent contamination of samples, materials containing Teflon were avoided during
188 whole treatment and analysis processes. All parts that had access to the samples were

189 cleaned using methanol and ultrapure water before use. Instrumental quantification
190 limits (IQLs) were defined as the lowest concentration of target compounds resulting
191 in a signal-to-noise (S/N) ratio of 3. Method quantification limits (MQLs) were defined
192 using a S/N ratio of 10. The IQLs and MQLs of all the target compounds are
193 summarized in Table S4.

194 The wastewater samples collected from the two WWTPs were analyzed separately to
195 avoid cross-contamination during instrumental analysis. Between every 10 samples
196 during sample treatment, one procedural blank using Milli-Q water was prepared by
197 following the same treatment procedures as described above, and the final
198 concentrations of these samples were subtracted from the levels in the blank samples.

199 A standard sample of 20 µg/L was added every 10-15 samples during the instrumental
200 injection in order to monitor instrumental stability (standard deviation less than ±10%).

201 The recoveries of most compounds in the wastewater ranged from 50-120% with
202 standard deviations below 20%. The average recoveries of the Internal Standards
203 ranged from 49% ± 17% ([¹³C₂]-PFDA) to 92% ± 19% ([¹⁸O₂]-PFHxS). Detailed
204 information of recoveries are listed in Table S5.

205 **Ratios of 6:2 Cl-PFESA to 8:2 Cl-PFESA**

206 The ratio of 6:2 Cl-PFESA to 8:2 Cl-PFESA expresses the variation of the two Cl-
207 PFESA's abundances relative to each other. For instance, increasing ratios could
208 indicate a selective process resulting in relative enrichment of either 6:2 Cl-PFESA.

209 The ratio, $R_{6:2/8:2}$, is given as:

210
$$R_{6:2/8:2} = \frac{C_{6:2 \text{ Cl-PFESA}}}{C_{8:2 \text{ Cl-PFESA}}} \quad (1)$$

211 C represents the concentration of the two Cl-PFESA.

212 **Results and discussion**

213 **Distribution and profile pattern**

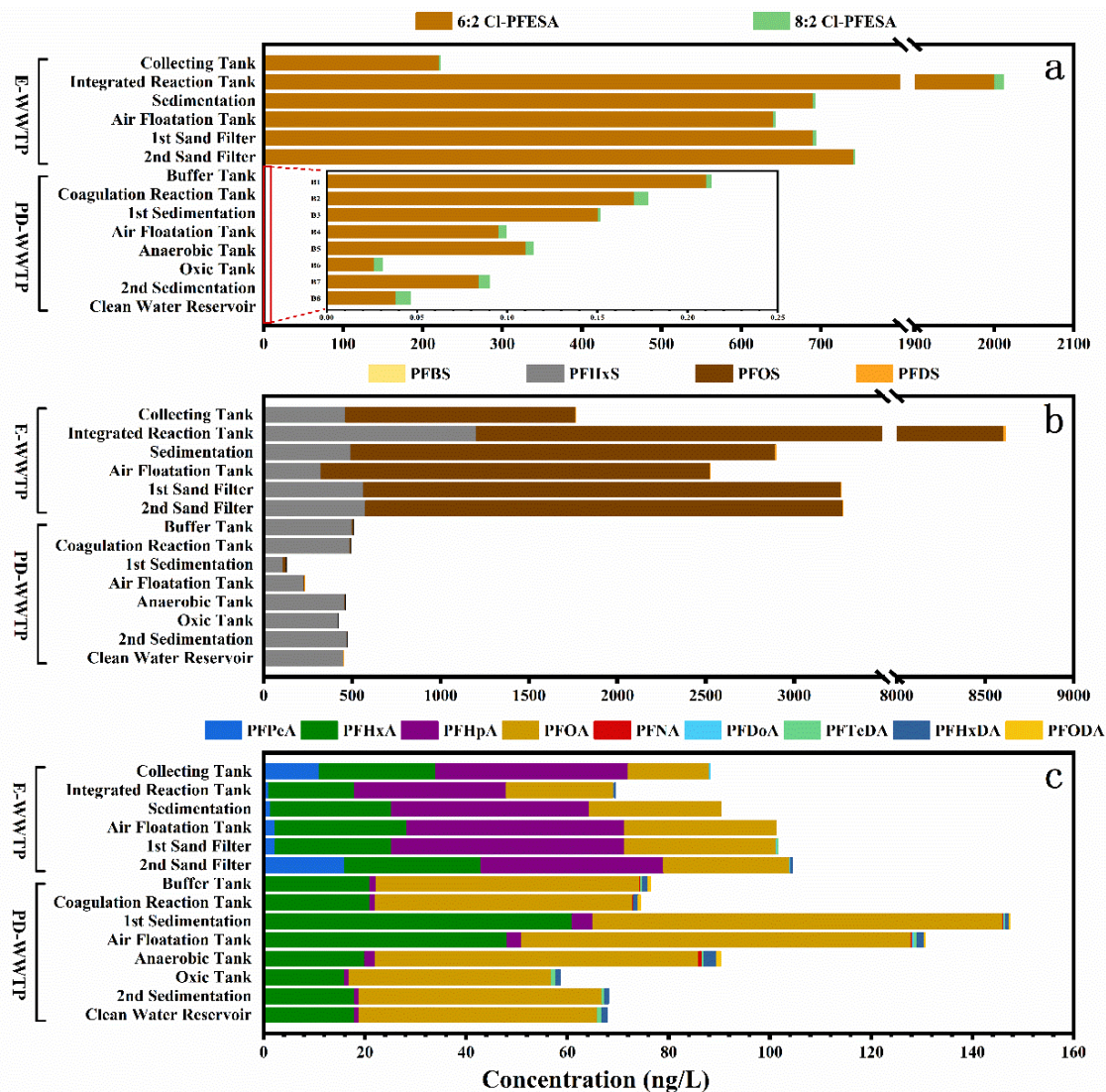
214 Wastewater samples were analyzed for Cl-PFESA and other target PFAS from the entire
215 treatment system of the two different WWTPs following the hydraulic retention time
216 (see Fig. 1). The aqueous concentrations of the target PFAS for the influent waters are
217 shown in Fig. 2. For the E-WWTP, 6:2 Cl-PFESA and 8:2 Cl-PFESA along with
218 seventeen different legacy PFAS were identified (see Fig. 2a). The total PFAS
219 concentration reached 2100 ± 130 ng/L in the influents and went up to 4200 ± 270 ng/L
220 in the effluents, indicating possible transformation and release from the precursors, or
221 variations in sorption/desorption processes, during the treatment processes (Table S6).
222 Concentrations of 6:2 Cl-PFESA and 8:2 Cl-PFESA in the E-WWTP influents were
223 220 ± 23 ng/L and 2.2 ± 0.49 ng/L. After water treatment these values went up to 740 ± 54
224 ng/L and 2.9 ± 0.37 ng/L, respectively (see Fig. 2a). Unlike PFCAs and PFSA, there are
225 no known precursors of Cl-PFSAs to the authors' knowledge. The major PFSA in the
226 wastewater, PFOS and PFHxS, occurred at average influent concentrations of 1300 ± 98
227 ng/L and 460 ± 18 ng/L, respectively, and at effluent concentrations of 2700 ± 190 ng/L
228 and 570 ± 37 ng/L, respectively (Fig. 2b). PFOS was the most abundant, accounting for
229 about 62% among the total PFAS in the wastewater, and the corresponding mean
230 concentrations was 1300 ± 98 ng/L, followed by 6:2 Cl-PFESA (220 ± 23 ng/L) and
231 PFHxS (460 ± 18 ng/L), accounting for 10% and 22%, respectively. Additionally,

232 PFCAAs including PFPeA, PFHxA, PFHpA and PFOA, contributed only about 4.2% to
233 the Σ PFAS concentrations in total (Fig. 2c).

234 Concerning the PD-WWTP, fifteen PFAS were identified and quantified. The Σ PFAS
235 concentration of the PD-WWTP influent reached 590 ± 39 ng/L, and this value was
236 slightly higher than the Σ PFAS concentration at the effluent (i.e. 520 ± 30 ng/L). Notably,
237 Cl-PFAES were also found in comparatively trace concentrations in the PD-WWTP,
238 with 6:2 Cl-PFAES being detected at 0.21 ± 0.03 ng/L in the influent and 0.04 ± 0.02
239 ng/L in the effluent, indicating PD-wastewater in this area might act as a novel, though
240 minor source for PFESAs environmental release. Given no evidence available for Cl-
241 PFAES applications in PD industries, we speculate that this is likely attributed to
242 "diffuse emissions" from small and unknown industries in this area. In general, PFAS
243 were inefficiently removed by applying the applied wastewater treatment techniques.
244 PFHxS, PFOA, PFHxA and PFOS were among the predominant species, with average
245 influent concentrations of 500 ± 36 ng/L, 52 ± 1.6 ng/L, 21 ± 0.94 ng/L and 8.2 ± 1.39 ng/L,
246 respectively, and effluent concentrations of 450 ± 28 ng/L, 47 ± 1.4 ng/L, 18 ± 1.7 ng/L and
247 2.2 ± 0.71 ng/L, respectively. In comparison, longer chain PFAS (> 8 carbon atoms)
248 were only observed at much lower concentrations (e.g. max < 1.12 ± 0.38 ng/L for
249 PFHxDA in effluent). As described in the Introduction, PFHxS are heavily applied in
250 the textile printing and dyeing process, and these occurred at 500 ± 36 ng/L, which
251 accounts for 85% mass of PFAS in the wastewater. PFOA, PFHxA and PFOS
252 contributed 8.8%, 3.6% and 1.4% to the Σ PFAS concentration, respectively.

253 In general, the composition of PFAS in the two industrial WWTPs are less

254 PFOA/PFOS-dominated comparing with the previous studies^{18, 20}. This is likely owing
255 to the fact that the short-chain and polyfluorinated PFAS were increasingly applied in
256 printing and dyeing as well as in electroplating industries^{22, 40}. For example, the chrome
257 mist suppressants used in electroplating industries have been most typically PFOS, but
258 now increasingly more alternative PFAS serve as active chemicals in chrome mist
259 suppressants formulations. Our results suggested that PFOS was predominant in the E-
260 wastewater (64%), and PFHxS and 6:2 Cl-PFESA were also abundant and account for
261 14% and 18%, respectively, indicating that these are being used increasingly.
262 Furthermore, the traditional textile PD-industry commonly uses fluorotelomer alcohols
263 (FTOHs) in their textile finishes, which are well-known precursors to PFCAs in textile
264 PD-wastewater²⁰. However, our study found that PFHxS (87%) was dominant PFAS,
265 and PFCAs only accounted for 13%. This might be due to increasing production and
266 application of perfluorohexane sulfonyl fluoride (PHxSF) as alternative for textile
267 surface treatment processes in China²⁵, which could degrade into PFHxS during
268 transformation processes^{25, 26}.



269

270 Figure 2. Concentrations of CI-PFESA and legacy PFAS in the wastewater of the two WWTPs. Panel (a)
 271 shows the concentration of 6:2 CI-PFESA and 8:2 CI-PFESA; Panel (b) shows the concentration PFASs;
 272 Panel (c) shows the concentration of PFCAs. The y-axis refers to the individual treatment steps for the
 273 two WWTPs, as listed in Figure 1.

274

275 **Mass flow analysis**

276 The mass flow of the individual PFAS in the two WWTPs on the day of sampling were
 277 derived (data available in Table S7). The mass flow of the major PFAS through the
 278 different treatment processes are shown in Fig. 3. Concerning the E-WWTP, the mass
 279 flow of total target PFAS were 2100 ± 130 mg/d in influents and went up to 4200 ± 270
 280 mg/d in effluents. The mass flow of 6:2 CI-PFESA and 8:2 CI-PFESA were observed

281 at 220 ± 23 mg/d and 2.2 ± 0.49 mg/d in the influents and 740 ± 54 mg/d and 2.9 ± 0.37
282 mg/d in the effluents (Fig. 3 (a1)). For other dominant species including PFOS and
283 PFHxS, the obtained corresponding values were 1300 ± 98 mg/d and 460 ± 18 mg/d
284 and in the influents 2700 ± 190 mg/d and 570 ± 37 mg/d in the effluents, respectively.
285 The mass flow in the effluent almost doubled compared to the influents. This could
286 indicate either large fluctuations in influents that our sampling campaign did not capture,
287 despite following the hydraulic flow of water, or alternatively variations in water
288 concentrations in the response to different treatment steps. A sharp change in the mass
289 flux was observed at the integrated reaction tank, after the "integrated reaction tank",
290 where all the PFASs and PFESAs increased significantly. Specifically, the mass flow
291 of 6:2 Cl-PFESA increased from 220 ± 23 mg/d (collection tank) to 2000 ± 18 mg/d
292 (integrated reaction tank), and after the same treatment procedures 8:2 Cl-PFESA
293 increased from 2.2 ± 0.49 mg/d to 12 ± 0.26 mg/d; PFHxS increased from 460 ± 18 mg/d
294 to 1200 ± 77 mg/d; PFOS increased from 1300 ± 98 mg/d to 7400 ± 3600 mg/d; PFDS
295 increased from 0.42 ± 0.14 mg/d to 12 ± 2.2 mg/d. We hypothesize that sharp increase of
296 pH by adding lime (pH=12) stimulated desorption of PFAS by deprotonating the anion
297 exchange sites on the suspended solids³⁸ causing the amount of freely-dissolved PFAS
298 to increase. After this initial spike in the reaction tank, 6:2 Cl-PFESA, 8:2 Cl-PFESA
299 and PFOS concentrations decrease in the sedimentation tank, due to losses to settling
300 sludge. The corresponding log Q_D values at the sedimentation step (log ratio of C_{sludge}
301 to C_{water}) are 4.5 and 6.0 for the 6:2 and 8:2 analogues in the E-WWTP and 3.7 and 5.2

302 in the PD-WTP, indicating strong sorption to sedimented sludge, particularly for the 8:2
303 analogue.

304 Comparing mass flows of Cl-PFESA to sludge and effluent leaving the facility gives
305 further insight.^{44, 45} The E-WWTP produces in the range of 2.3-3.0 tons sludge dw/day,
306 corresponding to removal of 50-66 g/day 6:2 Cl-PFESA and 7-10 g/day 8:2 Cl-PFESA
307 via sludge, much larger than the amount released as effluent, being 0.7 and 0.003 g/day,
308 respectively. Similarly, the PD-WWTP produces 0.60-0.75 tons dw/day, corresponding
309 to removal in sludge by 0.0004-0.0005 g/day 6:2 Cl-PFESA and circa 0.0002 g/day 8:2
310 Cl-PFESA, which is larger than the trace emissions from the effluent water of 5E-7 and
311 5E-9 g/day, respectively. This indicates that suspended solids are a substantial reservoir
312 for Cl-PFESA and other high sorbing PFAS. However, despite this removal via sludge,
313 the water concentration during sedimentation nevertheless remains elevated compared
314 to the influent concentration, indicating there still remains suspended solids that desorb
315 the target PFAS, and this concentration remains elevated through all subsequent
316 treatment steps (Figure 2).

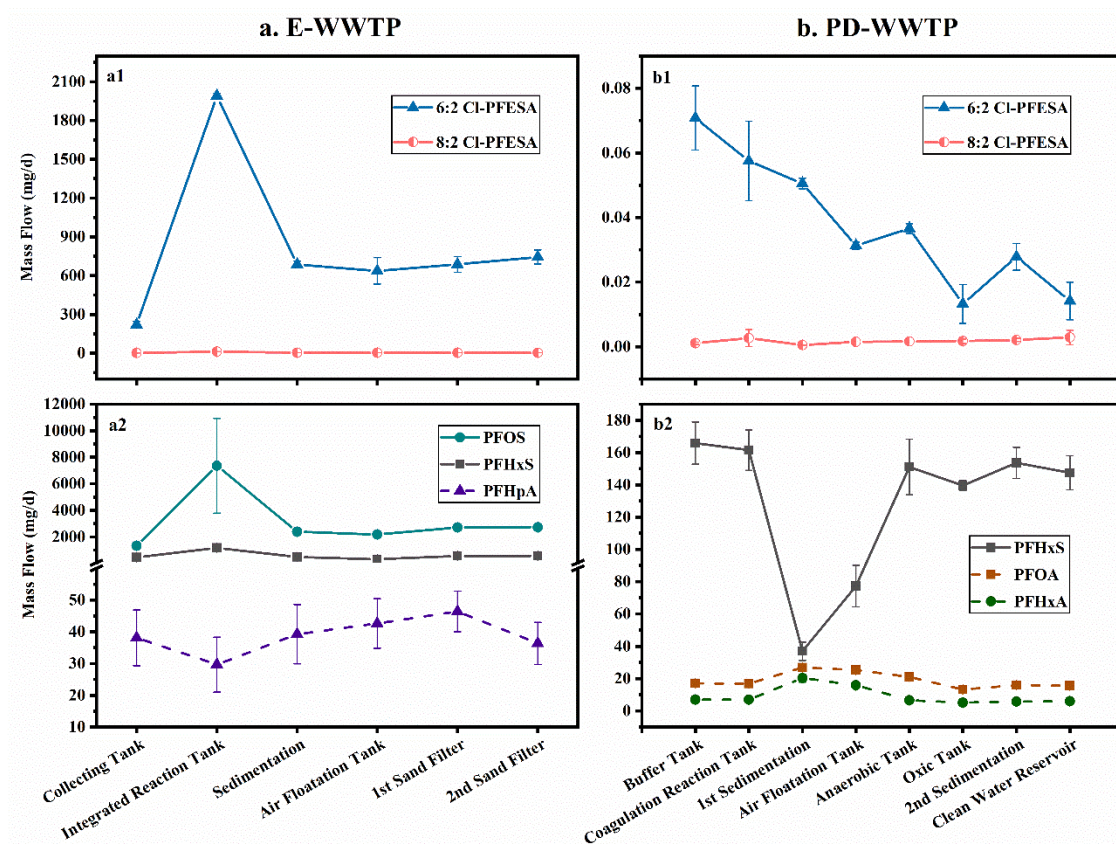
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318 However, other PFCAs did not show the same trend where PFPeA dropping from
319 11 ± 0.93 mg/d (collection tank) to 0.97 ± 0.2 mg/d (integrated reaction tank), PFHxA
320 showing little change from 23 ± 3.7 mg/d to 17 ± 0.28 mg/d and PFHpA showing little
321 change 38 ± 7.6 mg/d to 30 ± 8.7 mg/d for PFHpA. The different behavior for the PFCAs
322 may be due the carboxylates having a larger pKa than the sulfonates, and therefore less

323 impacted by the competition with hydroxyl radicals introduced by the pH increase by
324 adding lime^{38, 41}.

325 Concerning the PD-WWTP, the mass flow of total PFAS were 190 ± 13 mg/d in influents
326 and 170 ± 10 mg/d in effluents. Both 6:2 Cl-PFESA and 8:2 Cl-PFESA were detected in
327 the PD-WWTP at the sub nanogram-per-liter concentrations (see Fig. 3a), resulting in
328 corresponding mass flows of 0.069 ± 0.011 mg/d and 0.0011 ± 0.0003 mg/d at influents
329 (Fig. 3 (b1)), and different but still low concentrations at the effluents (0.013 ± 0.0063
330 mg/d and 0.0029 ± 0.0022 mg/d). In addition, the mass flow of the three predominant
331 compounds PFHxS, PFOA and PFHxA in the influents reached 170 ± 12 mg/d, $17 \pm$
332 0.52 mg/d and 6.9 ± 0.31 mg/d, respectively. There are only small differences on the
333 values obtained in the effluents (i.e. 150 ± 9.2 mg/d, 16 ± 0.46 mg/d, and 6.1 ± 0.57
334 mg/d, respectively). However, the mass flow fluctuated during the treatment processes.
335 For instance, a dramatic change occurred at after the first sedimentation, where PFHxS
336 mass flow decreased from 160 ± 10 mg/d to 37 ± 4.6 mg/d, indicating it was removed
337 by sludge. However, PFOA and PFHxA presented an increasing trend, with increasing
338 PFOA mass loads from 17 ± 0.3 mg/d (reaction tank) to 27 ± 1.8 mg/d (first
339 sedimentation), and from 7 ± 0.43 mg/d (reaction tank) to 20 ± 2 mg/d (first
340 sedimentation) for PFHxA. This could be due to precursor transformation, as the textile
341 PD industry in China is known to emit several PFCA precursors, specifically
342 fluorotelomer alcohols^{42, 43}. The individual PFHxS mass flux then went back to their
343 original levels after air floatation and after anaerobic digestion (see Fig. 3); hence, this
344 sudden dip after sedimentation might be due to selective precipitation/aggregation of

345 PFHxS³⁵, and followed by yield of PFHxS through degradation of perfluorohexane
 346 sulfonyl fluoride (PFHxS)-based derivatives during anaerobic tank treatment.



347
 348 Figure 3. Mass flow of the CI-PFESA and predominant PFAS (mg/day) obtained in different treatment
 349 steps in the two WWTPs. Panel (a) shows the data for the E-WWTP, and Panel (b) shows the data for the
 350 PD-WWTP. The dashed lines represent PFAS with carboxylic acids (PFCAs), and the solid lines
 351 represent PFAS with sulfonic acids (PFSAs and CI-PFESAs).

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 354

355 Fate behavior of 6:2 and 8:2 CI-PFESA

356 The fate of 6:2 CI-PFESA and 8:2 CI-PFESA in wastewater during treatment processes
 357 of the two WWTPs were compared by calculating $R_{6:2/8:2}$ at each stage, as summarized
 358 in Fig. 4. An increase in this ratio represents 6:2 CI-PFESA-enrichment processes due
 359 to preferential desorption of 6:2 CI-PFESA or preferential sorption of 8:2 CI-PFESA.
 360 Trends in such (de)sorption are also evident from the PFAS removal efficiency at each

361 step of the treatment process for 6:2 and 8:2 Cl-PFESA, as presented in Table S10.

362 For the E-WWTP, $R_{6:2/8:2}$ initiated at 100 and went up to 260 for the entire treatment
363 processes, indicating a much higher relative abundance of 6:2 Cl-PFESA (>90%) in
364 both influents and effluents. The corresponding removal efficiency of 6:2 Cl-PFESA
365 and 8:2 Cl-PFESA were -230% and -32%, respectively (where negative values imply
366 an increase in concentration, see Table S10), implying net-desorption from suspended
367 solids with more occurring for 6:2 Cl-PFESA. This net desorption largely occurred
368 from the first aeration step, with a 8-fold vs 4-fold increase in 6:2 and 8:2 Cl-PFESA,
369 respectively. The $R_{6:2/8:2}$ increased most substantially afterafter sedimentation treatment,
370 to a value of 230. This is possibly due to preferential removal of 8:2 Cl-PFESA (73%
371 removed) compared with 6:2 Cl-PFESA (66% removed).

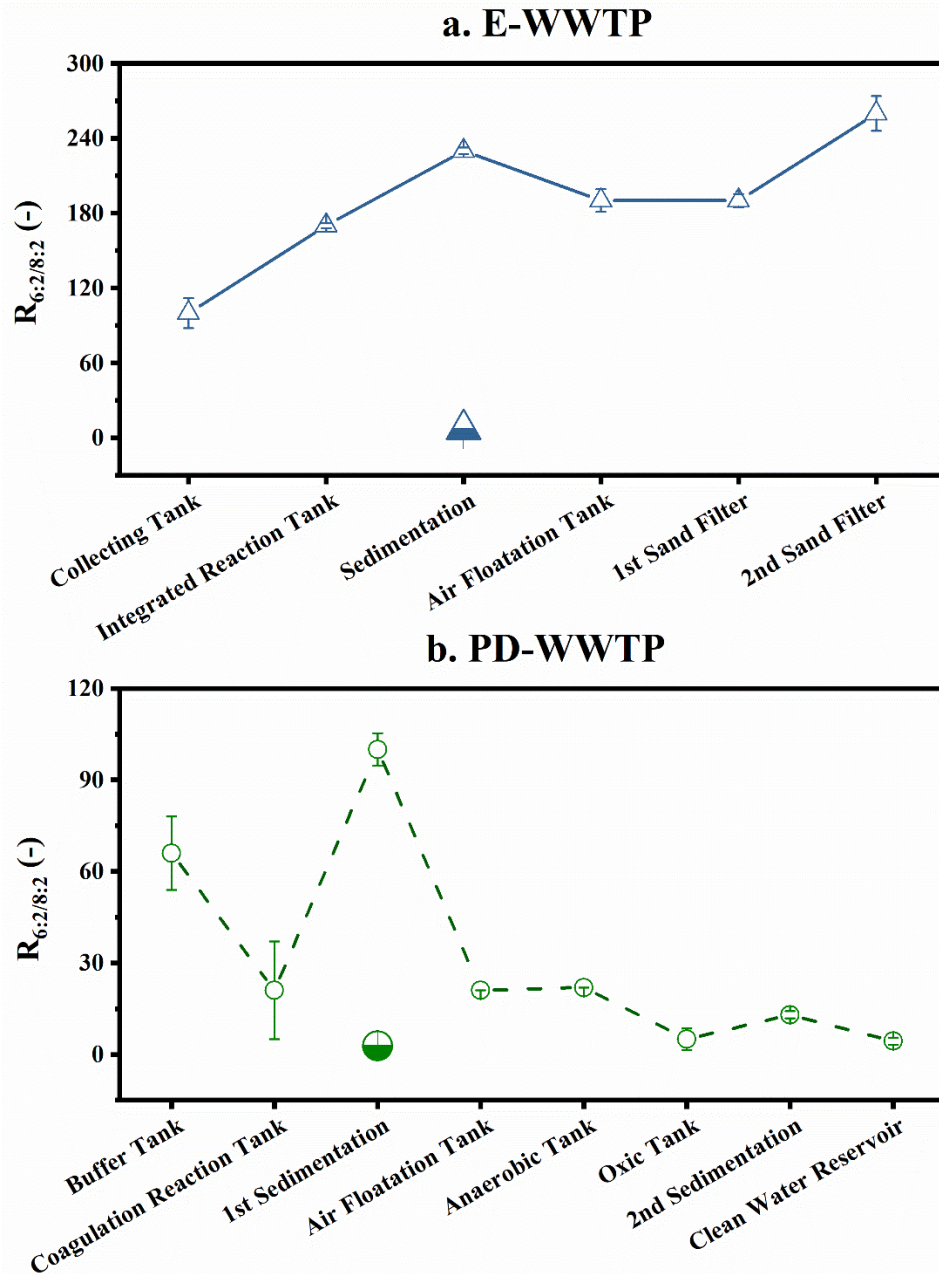
372 For the PD-WWTP, $R_{6:2/8:2}$ at the influent was 66 ± 12 different from the value obtained
373 at the influent of E-WWTP at 100 ± 12 . The $R_{6:2/8:2}$ ratio then went down to 7 after entire
374 PD-wastewater treatment processes. This enrichment of the lesser sorbing 8:2 Cl-
375 PFESA seems counter-intuitive, and mainly occurred during the "air flotation" step,
376 which saw an increase in 8:2 Cl-PFESA, yet a decrease in 6:2 Cl-PFESA; a likely
377 explanation for this is the $R_{6:2/8:2}$ for the PD-PFES is much more prone to analytical
378 errors, as the concentrations are quite near the quantification limit, particularly the 8:2
379 Cl-PFESA (Table S6).

380 Considering the sludge fraction, the concentrations of 6:2 Cl-PFESA and 8:2 Cl-
381 PFESA in the sludge are 22000 ng/g and 3200 ng/g (E-WWTP) and 0.72 ng/g and 0.26
382 ng/g (PD-WWTP), respectively (Table S13). The corresponding 6:2/8:2 Cl-PFESA

383 ratios obtained from the E-WWTP and PD-WWTP are 6.9 ± 0.04 and 2.8 ± 0.16 ,
384 respectively, which can be compared to the corresponding water fraction at 230 ± 2.7
385 and 100 ± 5.3 , respectively. A dip of the ratios in the sludge indicates enrichment of 8:2
386 Cl-PFESA in the sludge during sedimentation, due to its strong sorption interactions
387 described above. A similar trend was observed in a previous study, where a much
388 smaller 6:2/8:2 Cl-PFESA ratios were obtained from the sludge in the municipal
389 WWTP compared to the ratios in the original product solutions.³⁸

390 As presented above, the loss in Cl-PFESA towards the sludge fraction were substantial,
391 but the impact on the water concentrations minimal. We hypothesize from these results
392 that one underlying process causing fluctuation of Cl-PFESA flux in the water could be
393 related to remaining suspended solids to be a substantial source of Cl-PFESA, where
394 metal ions released from activated sludge or hydroxyl radicals from the lime could lead
395 to decrease sorption of the Cl-PFESA due to competitive sorption at anionic exchange
396 sites.^{38, 41}

397 The temporary removal efficiencies of 6:2 Cl-PFESA and 8:2 Cl-PFESA in the waste
398 water phase during sedimentation tank treatment in E-WWTPs were 66% and 73%; 10%
399 and 81% for PD-WWTPs, respectively, are worth note that these were not retained or
400 ultimately effective in subsequent water filtration steps.



401

402 Figure 4. Variation of 6:2 Cl-PFESA over 8:2 Cl-PFESA ratios ($R_{6:2/8:2}$, see Eq. 1) during individual

403 wastewater treatment processes in E-WWTP (panel a) and PD-WWTP (panel b). The filled symbols

404 represent the 6:2/8:2 Cl-PFESA ratios obtained from wastewater samples; the semi-filled symbols

405 show the ratios obtained from sludge samples. The ratio is expected to increase in the case of

406 preferential sorption of 8:2 Cl-PFESA.

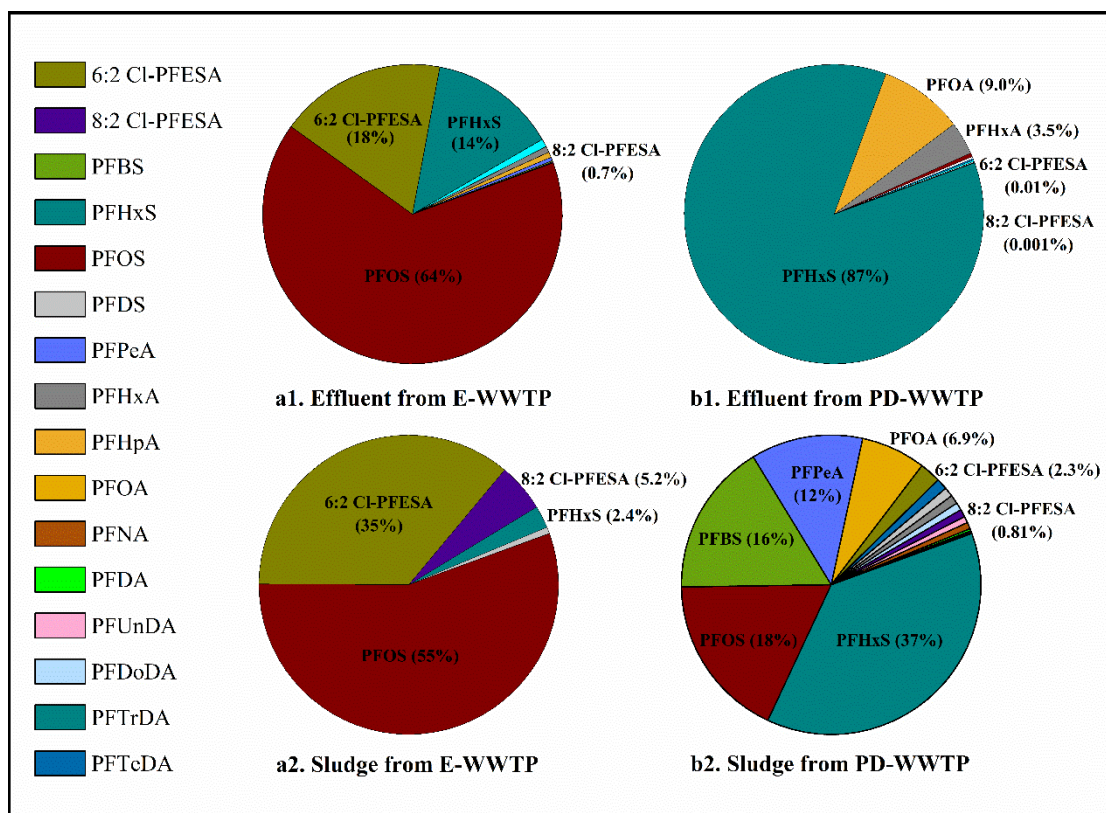
407

408 **Environmental implications**

409 PFAS were intensively applied at the two industrial sites, which then emitted PFAS
410 towards the wastewater treatment plants. According to the PFAS emission inventories
411 obtained in the effluents (see Fig. 5), PFASs were predominant in both WWTPs. Both
412 6:2 Cl-PFESA and 8:2 Cl-PFESA were found in the effluent and sludge of E-WWTP at
413 high abundances due to their extensive applications as chrome mist suppressants in
414 China, but also appeared in trace concentrations in the PD-WWTP likely due to diffuse
415 emission sources. We hypothesize that desorption of Cl-PFESAs from suspended solids,
416 to which they are substantially sorbed, could have occurred during the addition of high
417 pH lime and flocculants (e.g. PAM) in the reaction tank, causing the 6:2 Cl-PFESA and
418 8:2 Cl-PFESA mass flux to increase in the water phase; testing for this could be the
419 focus of a future study to investigate this hypothesis. A large amount of PFAS-
420 containing wet solid wastes were produced by the two investigated WWTPs. In the
421 present case, these industrial sludges are sent to incineration.

422 Future work should focus on characterization of loss processes from sludge
423 sedimentation based on dynamic colloidal/suspended particle sorption behavior (e.g. as
424 observed in the reaction tank), in particular, in presence of metal ions, flocculants and
425 pH. Recent advances on non-target screening tools based on high-resolution mass
426 spectrometry could help to identify novel homologue series of n:2 Cl-PFESAs and their
427 degradation intermediates in wastewater and impacted environments^{33, 46}. This would
428 also add valuable information to identify local PFAS hotspots as well as to complete
429 emission inventories of different PFAS-relevant manufacturers in China.

430 CI-PFESA contaminated industrial wastewaters were not efficiently remediated by the
 431 WWTPs investigated, even though much was captured by the sludge. Specially, both
 432 6:2 CI-PFESA and 8:2 CI-PFESA concentrations even increased in the effluents of E-
 433 WWTPs. So far, only advanced and relative expensive water treatment techniques
 434 including activated carbon filtration, anion exchange methods, and advanced
 435 oxidation/reduction processes, provide solutions for removal of some PFAS⁴⁷⁻⁵¹, but
 436 their efficiency seems to decrease the smaller the size of the PFAS. Currently, only non-
 437 PFAS substitutes together with innovative, expensive remediation techniques are the
 438 only options for the reduction PFAS and fluorinated alternatives to them. Innovation
 439 towards alternatives to PFAS is recommended for both the textile printing and dyeing
 440 industry, as well as the electroplating industry.



441
 442 Figure 5. Alternative and legacy PFAS fraction (%) in the effluents and the sludge collected from the two

443 WWTPs. The pie chart on the left represents the E-WWTP (a1; a2), and the chart on the right represents
444 the PD-WWTP (b1; b2).

445 **Supporting Information**

446 The supporting information is available free of charge on the ACS publication website
447 at DOI: XXXXX.

448 Description of calculations for mass flow and removal efficiency; instrumental
449 parameters and relevant information of wastewater treatment plants and target analytes
450 (Table S1-S5); data evaluation of PFAS compositions and dynamics during treatment
451 processes (Table S6-S10, S13; Figure S1); pH values at each treatment step (Table S11-
452 S12); Content (ng/g) of PFASs in the sludge samples collected in the two WWTPs
453 (Table S13).

454

455 **Acknowledgments**

456 B.J. acknowledges support from Guangdong Foundation for Science and Technology
457 Research (2020B1212060053; 2019A1515011035), and a grant from State Key
458 Laboratory of Organic Geochemistry, Chinese Academy of Sciences (SKLOG2020-4).

459 G.Z. acknowledges funding from Guangdong Science and Technology Projects
460 (2018B030324002). H.P.H.A. acknowledges the Research Council of Norway
461 SLUDGEFFECT (302371/E10). The authors thank the three anonymous reviewers and
462 Dr. Zhen Zhao (Shanghai Ocean University) for their constructive comments and
463 advice.

464

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